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8-31-99 V. Hughes
 Date August 31, 1999
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Sir: This is a request for filing a
 (X) Continuation application,
 () Divisional application,

under 37 CFR §1.60, of pending prior application Serial No. 08/934,367 filed on September 19, 1997 of Needleman et al. for AN IMMUNOLOGICAL PROCESS AND CONSTRUCTS FOR INCREASING THE HDL CHOLESTEROL CONCENTRATION BY DNA VACCINATION

1. (X) Enclosed is a copy of the prior application, including the oath or declaration as originally filed. The attached application papers are a true copy of prior application Serial No. 08/934,367 as originally filed on September 19, 1997, except that the express mail certification present at the top of page 1 of the parent application has been replaced by a similar certification reciting the filing date and no amendments referred to in the oath or declaration as originally filed introduced new matter. This statement is made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

2. (X) The filing fee is calculated below:

CLAIMS AS FILED IN THE PRIOR APPLICATION, LESS ANY CLAIMS
 CANCELLED BY AMENDMENT BELOW

For	No. Filed	No. Extra	Rate	Fee
Basic Fee. . . .				\$ 760.00
Independent Claims . . .	1 - 3 =	0	x \$82.00 =	\$ _____ 0
Total Claims	8 - 20 =	0	x \$18.00 =	\$ _____ 0
		Total Filing Fee		\$ _____ 760.00
Statement of Status as Small Entity Reducing Filing Fee by Half to				\$ _____ .00

3. () Verified statements (Declarations) claiming small entity status were filed in the prior application and small entity status for this application is proper and desired.
4. (X) A check in the amount of \$ 760.00 is enclosed.
5. (X) The Commissioner is hereby authorized to charge any additional fees which may be required for this application under 37 CFR §§1.16-1.17, or credit any overpayment, to Deposit Account No. 23-0920. A duplicate copy of this sheet is enclosed.
6. (X) Cancel in this application original claims 1, 2, 9-11, and 14-31 of the prior application before calculating the filing fee.
7. (X) Amend the specification by inserting before the first line the sentence: --This is a (X) continuation, () division, of application Serial No. 08/934,367, filed September 19, 1997.--
8. () Transfer the drawings from the prior application to this application and abandon said prior application as of the filing date accorded this application. A duplicate copy of this sheet is enclosed for filing in the prior application file. (May be used only if signed by person authorized by §1.138 and before payment of base issue fee.)
- 8a. () New formal drawings are enclosed.
9. (X) Other: STATEMENT UNDER 37 CFR § 1.821(f) and computer readable disk with prior sequence listing from parent application
10. () Priority of application Serial No. _____ filed on _____ in _____ is claimed under 35 USC §119.
11. (X) The prior application is assigned of record to Monsanto Company.
12. (X) The power of attorney in the prior application is to :

Edward P. Gamson Registration No. 29,381
WELSH & KATZ, LTD.
120 South Riverside Plaza
22nd Floor
Chicago, Illinois 60606
and other members of the firm.

- (a) ☒ The power appears in the original papers in the prior application.
- (b) ☐ Since the power does not appear in the original papers, a copy of the power in the prior application is enclosed.
- (c) ☒ Address all future communications to:
WELSH & KATZ, LTD.
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Telephone: (312) 655-1500
(May be completed only by applicant, or the attorney or agent of record.)
- (d) ☒ The executed Power of Attorney document filed in the prior application.
- (e) ☐ The executed Verified Statements claiming small entity, filed in the prior application.

August 31, 1999



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AN IMMUNOLOGICAL PROCESS AND CONSTRUCTS
FOR INCREASING THE HDL CHOLESTEROL
CONCENTRATION BY DNA VACCINATION

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Description

Cross-Reference to Related Applications

This is a continuation-in-part of application
10 Serial Nos. 08/785,997 and 08/788,882, both filed
January 21, 1997.

Technical Field

The present invention relates to a process for
15 inducing antibodies in a mammal by immunization with DNA
that encodes cholesteryl ester transfer protein (CETP)
or a portion thereof, and more particularly to an
immunological process for ameliorating
dyslipoproteinemias in immunized mammals characterized
20 by low HDL/LDL cholesterol ratios by means of those
induced antibodies as well as specific DNA constructs
for use in that process.

Background of the Invention

25 Cholesteryl ester transfer protein (CETP) is
an acidic plasma glycoprotein that plays a critical role
in establishing high density lipoprotein (HDL), low
density lipoprotein (LDL), and very low density
lipoprotein (VLDL) cholesterol blood plasma levels and
30 lipid composition in plasma [L. Lagrost, *Biochem.*
Biophys. Acta., 1215:209-236 (1994)]. Several studies,
some of which are discussed below, have demonstrated
that CETP mediates the transfer of cholesteryl esters
(CE) from HDL particles to LDL and VLDL particles, as
35 well as mediating the transfer of triglycerides (TG)

from LDL and VLDL to HDL particles. This reciprocal exchange of CE and TG is the primary means of providing CE to LDL and VLDL particles in many mammalian species. CETP thus mediates the balanced exchange of cholesteryl esters (CE) and triglycerides (TG) between pro-atherogenic (LDL and VLDL) and anti-atherogenic (HDL) lipoprotein fractions.

Mammalian species whose blood plasma contains CETP such as humans and other primates, rabbits, and hamsters suffer atherosclerosis and heart disease when exposed to diets rich in cholesterol. Other animal species such as mice, rats and dogs lack plasma CETP (measured as transfer activity) and are not susceptible to dietary cholesterol-induced atherosclerosis.

That CETP contributes to the pathogenesis of atherosclerosis in humans has been strongly supported by transgenic mouse studies [G. Melchior et al., *Trends in Card. Med*, 5:83-87 (1995)]. For example, transgenic mice having a mini gene of cynomolgus monkey CETP cDNA plus the proximal region of the CETP promoter show dietary cholesterol regulation of CETP levels similar to those seen in humans, hamsters and monkeys. Those transgenic mice expressing high levels of the monkey CETP (levels comparable to human dyslipidemias) exhibit: increased LDL+VLDL cholesterol and apo-B and, decreased HDL cholesterol, LDL-receptor and HMG-CoA reductase mRNA. Atheroma could be induced by high fat diet in transgenic mice with the cynomolgus monkey CETP transgene.

The CETP amino acid residue and nucleotide sequences of mammalian species have been characterized. For example, the human CETP DNA sequence of SEQ ID NO:1 has been determined [D. Drayna et al., *Nature*, 327:632-634 (1987)]. The rabbit CETP DNA sequence of

SEQ ID NO:27 has also been characterized [M. Nagashima
et al., *J. Lipid Res.*, 29:1643-1469 (1988); Kotake et
al., *J. Lipid Res.*, 37:599-605(1996) and Kotake et al.,
Biochim. Biophys. Acta., 1347:69-74 (1997)], as has the
5 cynomolgus monkey CETP sequence SEQ ID NO:31 [M. E. Pape
et al., *Atherosclerosis and Thrombosis*, 11:1759-1771
(1991)]. The human CETP protein is 476 amino acid
residues long, whereas the rabbit CETP protein is 496
amino acid residues long, and the cynomolgus monkey
10 sequence contains 476 residues.

CETP may be a key factor for the global
regulation of atherogenicity of plasma lipoproteins in
patients with atherosclerosis or coronary artery disease
(CAD). CAD is the number one cause of morbidity and
15 mortality in western society. Patients at increased
risk for developing coronary artery disease typically
exhibit an enhanced level of CETP activity. It has also
been reported that CETP has higher affinity for oxidized
LDL than native LDL molecules [L. Lagrost, *Biochem.*
20 *Biophys. Acta.*, 1215:209-236 (1994)]. High levels of
LDL cholesterol (>180 mg/dl) [*J. Am. Med. Assoc.*,
269:3015-3023 (1993) and A. L. Gould et al.,
Circulation, 91:2274-2282 (1995)]; and low levels of HDL
cholesterol (<35 mg/dl) [G. Assman et al., *Excerpta*
25 *Medica*, 46-59 (1989) and V. Manninen et al.,
Circulation, 85:37-45 (1992)] have been reported to be
important contributors to the development of
atherosclerosis.

Individuals who possess genetic deficiencies
30 of the CETP protein have elevated HDL cholesterol
levels. Heterozygotes have HDL levels 15-20 percent
above non-affected controls. It has been suggested that
there is a 2-3 percent decrease in incidence for CAD for
each 1 mg/dl increase in HDL cholesterol after

correction for other risk factors [D. J. Grodon et al., *Nature*, 79:8-15, (1989)].

5 In an experimental model of CETP deficiency in hamsters, it has been shown that passive transfer of mouse anti-human CETP monoclonal antibodies (1C4) inhibited hamster plasma CETP CE transfer by 70-80 percent at all times up to 24 hours following injection of 500 μ g of 1C4 (approximately 3.7 mg/kg body weight). That inhibition of hamster CETP-mediated transfer *in* vivo increased hamster HDL cholesterol by 33 percent, increased HDL-CE by 31 percent and decreased HDL-TG by 42 percent. These results indicate an example of mammalian (hamster) CETP-mediated CE-TG exchange being disrupted by xenogeneic anti-human CETP monoclonal antibodies, and further demonstrate the use of hamsters as a pre-clinical model for testing CETP inhibition [B. J. Gaynor et al., *Atherosclerosis*, 110(1):101-109 (1994)].

20 In another study reported by G. W. Melchior et al., *J. Biol. Chem.*, 270(36):21068-74 (1995) cynomolgus monkey CETP was shown to have two neutral lipid binding sites. A monoclonal antibody to purified cynomolgus monkey CETP identified as CMTP-2 was capable of severely inhibiting triglyceride (TG) transfer, but had a variable effect on cholesteryl ester (CE) transfer.

25 Thus, when the monoclonal antibody was administered sub-cutaneously to cynomolgus monkeys at a dose that inhibited TG transfer in the plasma by more than 90 percent, there was no detectable effect on the high density lipoprotein cholesterol level, but the HDL-TG levels decreased from 13 to 1 mol/mol of HDL. A Fab antibody fragment had no effect on CE transfer, but completely blocked TG transfer. Another type of inhibitor, 6-chloromercuric cholesterol, severely

inhibited CE transfer with minimal inhibition of TG transfer. When both the inhibitory monoclonal antibody and the 6-chloromercuric cholesterol were added to the assay, both CE and TG transfer were inhibited, indicating that the inhibitors did not compete for the same binding site on CETP. This study indicated that *in vivo* administration of xenogeneic monoclonal antibodies uncoupled CE and TG transfer.

The inhibitory effects of anti-sense RNA on expression of CETP protein were reported using vaccinia virus as an expression system. [M. H. Lee et al., *J. Biochem. Mol. Biol.*, 28(3):243-248 (1995)]. The cDNA from CETP was inserted into a transfer vector (pSC11) in sense and anti-sense orientations and then used to construct recombinant vaccinia viruses. Decreased expression of the exogenous CETP cDNA in mouse cells was clearly evident in the Northern and Western blot analyses as the dose of anti-sense expression increased. Also, in the CETP assay, the CETP activity was decreased compared to the activity obtained from the cell extracts infected with sense constructs only.

More recently, Sugano et al., *J. Biol. Chem.*, 271(32):19080-19083 (1996) reported upon the *in vivo* effects of anti-sense CETP RNA administration to rabbits. In that report, decreases in total cholesterol and CETP activity levels were found 24, 48 and 96 hours following anti-sense CETP administration, as was an increase in plasma HDL cholesterol at 48 hours.

Other methods of inhibition of CETP-mediated transfer are described in the literature. For example, data from Parke-Davis company has shown that infusion of 10 to 20 mpk of the small molecule compound referred to as PD 140195 into rabbits inhibited CETP activity within 30 minutes (measured in an *ex vivo* assay) [C. Bisgaier

et al., *Lipids*, 811-818 (1994)]. Schering-Plough Company has published on the isolation of Wiedendiol-A and -B from a marine sponge and has shown that this class of compounds to be low μ M inhibitors of CETP-mediated CE transfer *in vitro* [S. Coval et al., *Bioorganic & Med. Claim. Lett.*, 5:605-619 (1995)].

Currently, nicotinic acid and the fibrate drugs are the only small molecule drug therapies that cause significant rises in HDL cholesterol. These drugs are poorly tolerated and must be taken daily. Therapeutic doses of these drugs lead to 15-20 percent increases in HDL cholesterol.

Three mouse monoclonal antibodies to human CETP that recognize a similar epitope on CETP, caused parallel and complete *in vitro* immunotitration of human plasma CE and triglyceride transfer activities, but only partial inhibition of phospholipid transfer activity [C. B. Hesler et al., *J. Biol. Chem.*, 263(11):5020-5023 (1988)]. Those three monoclonals were originally designated 5C7, 2H4 and 7E1, but in more recent publications of the authors, those monoclonals are referred to as TP2, TP1 and TP3, respectively.

Monoclonal antibody TP2 is directed against an epitope within the last 26 amino acids of CETP (SEQ ID NO:29) [T.L. Swenson et al., *J. Biol. Chem.*, 264:14318-14326 (1989)], and more particularly to an epitope between about positions 465 and 475 of SEQ ID NO:28 [Tall, *J. Lipid Res.*, 34:1255-1274 (1993)]. That monoclonal has been shown to block CETP-mediated lipid transfer by limiting access to lipid-binding sites in the carboxy-terminus of CETP.

In an *in vivo* study using the xenogeneic mouse monoclonal antibodies (TP1) to CETP, rabbits were intravenously injected with TP1, or irrelevant

monoclonal antibodies or saline (control), resulting in an initial 70 percent inhibition of CETP-mediated CE transfer activity. Inhibition was 45 percent after 48 hours for the TP1-injected animals. HDL-CE increased in TP1-treated animals and reached levels that doubled over initial and control values at 48 hours. HDL-TG fell reciprocally, but HDL protein did not change, suggesting a CE for TG exchange. VLDL CE-TG ratio also decreased. CETP inhibition delayed the initial clearance of radioactively-tagged HDL, suggesting that CETP plays a quantitative role in HDL-CE catabolism in the rabbit, promoting the exchange of TG for CE, and the clearance of CE from plasma [M. E. Whitlock et al., *J. Clin. Invest.*, 84:129-137 (1989)].

In further animal studies with hamsters, a single sub-cutaneous injection of TP2 monoclonal antibodies in another illustration of passive administration of xenogeneic antibodies decreased CETP-mediated activity by 58 percent, lowered LDL+VLDL cholesterol 32 percent and raised HDL cholesterol 24 percent [G. Evans et al., *J. Lipid Res.*, 35:1634-1645 (1994) and S. Zuckerman et al., *Lipids*, 30:307-311 (1995)]. The effect of the TP2 monoclonal antibodies on CETP-mediated CE transfer inhibition was evident within 24 hours after injection and was maximized by 4 days. Lipoproteins returned to control levels 14 days after TP2 administration. The shift in the ratio of VLDL+LDL cholesterol to HDL cholesterol levels due to TP2 monoclonal antibody administration was more significant in hypercholesterolemic hamsters.

TP2 also has a higher efficacy in hamsters fed with a western diet enriched in cholesterol. CETP-mediated activity was reportedly increased in these animals 2-fold over chow-fed hamsters.

The preparation of recombinant CETP molecules has been reported by several research groups. For example, in a study reported recently, glutathione S-transferase-human CETP fusion protein (86 kDa) was expressed using vaccinia viral transfer vectors transfected into CV-1 monkey kidney cells. Using a Western blot assay, the fusion protein was identified by polyclonal antibodies against the carboxy-terminal active region of CETP fused with GST. After cleavage of the GST portion of the fusion protein, the purified CETP showed biological activity in a CETP *in vitro* assay [W. H. Yoon et al., *Mol. Cells*, 5(2):107-113 (1995)] and M. K. Jang et al., *J. Biochem. Mol. Biol.*, 28(3):216-220 (1995)].

It has also been reported that specific rabbit polyclonal antibodies were produced by immunization with a GST-CETP fusion protein. A full-length CETP cDNA clone isolated from a human heart λ gt11 library was used to provide the C-terminal 94 bp of CETP after a full length CETP molecule expressed in *E. coli* was found to be insoluble. The λ gt11 cDNA library was subcloned into pGEX plasmid and a GST-CETP fusion protein was expressed in *E. coli*. The CETP-GST fusion protein was purified by glutathione-Sepharose-4B affinity chromatography and used as an antigen for the production of rabbit polyclonal antibodies. The antibodies showed good titers, not only against the GST-CETP fusion protein, but also against a mixture of synthetic peptides corresponding in sequence to two 16-mers from the carboxy-terminal region of human CETP. The antibodies were said to be useful as an immunological tool for a CETP assay [N.W. Jeong et al., *Mol. Cells*, 4(4):529-533 (1994)].

To date there are no published reports on the long-term inhibition of CETP-mediated CE transfer. Passive immunization with the use of xenogeneic antibodies can only be utilized for a short-term period of time because host animals develop antibodies to the xenogeneic immunoglobulin.

One strategy for the generation of a host-induced immune response as compared to xenogeneic antibody use is based on DNA vaccine or "genetic immunization" technology. [W.M. McDonnell et al., *N. Engl. J. Med.*, 334(1): 42045 (1996).] Typically, a DNA vaccine contains a vector that includes one or more genes encoding an antigenic portion of a virus, such as an envelope, surface or a core protein. Host cells take up the DNA vector, express the heterologous gene, and produce the corresponding viral protein inside the cell.

One advantage of this approach is that the viral protein enters and is processed by the cell's major histocompatibility complex (MHC) class I pathway. MHC class I molecules carry peptide fragments to the cell surface where they evoke cell-mediated immunity by stimulating CD⁺8 cytotoxic T cells. Standard vaccine antigens enter cells by phagocytosis or endocytosis and are processed through the MHC class II system, to stimulate antibody responses.

The rapid development of DNA vaccine technology within the past several years was spurred by the report that direct injections of a gene from the influenza A virus could be used to immunize mice against the disease. [J.B. Ulmer et al., *Science*, 259: 1745-1749 (1993).] Since then, induction of antibodies has been reported for a variety of pathogen-derived proteins including the influenza NP, HA, M1 proteins; HIV Env, Gag, Rev proteins; bovine herpes virus gp; hepatitis B

virus surface and core antigens; rabies virus gp, NP, Plasmodium sp. CSP; Leishmania major gp63; Mycobacterium tuberculosis HSP65, Ag85; Hepatitis C virus nucleocapsid protein; Herpes simplex virus gB, gD, ICP27; Papillomavirus L1; Human T-cell leukemia virus type 1 Env; Lymphocytic choriomeningitis virus NP; Bacillus thuringiensis Endotoxin; Mycoplasma pulmonis ND; and Salmonella typhi OmpC porin. [See the references cited in J.B. Ulmer et al., *ASM News*, 62(9): 476-479 (1996).] Induction of cytotoxic T lymphocytes (CTL), protective immunization, or both, has been reported for each of these examples.

The potential advantages of DNA vaccines, in terms of efficacy and cost, over vaccines prepared by traditional approaches is considered to be as significant as two other major advances that were developed over the past century. The first development pioneered by Louis Pasteur was the use of attenuated and killed forms of microorganisms. The second major development was the use of defined components of whole organisms and the use of purified recombinant proteins. The DNA vaccine approach, has been termed the "third vaccine revolution." [B. Dixon, *Bio/Technology*, 13(5): 420 (1995).]

DNA vaccines offer several advantages over other types of vaccines. [J.B. Ulmer et al., *ASM News*, 62(9): 476-479 (1996).] First, expression of antigen encoded on the vector introduced into host cells leads to production of structurally relevant proteins, which are appropriately modified, and induction of cytotoxic T lymphocytes. Second, DNA vaccines induce CTL responses without resorting to complicated protein formulations or to attenuated live organisms. Attenuated live organisms, such as bacteria, or viruses, also have a

greater inherent ability to mutate to more virulent forms. Third, expression of antigens after DNA vaccination can persist, for months in some cases, sufficient to promote the induction of memory immune
5 cells. Still further, DNA vaccines can easily be injected intramuscularly or intradermally in simple aqueous solutions, or coated onto metal particles which are blasted into cells with gene guns, which facilitates administration and their subsequent analysis. [Finan,
10 E. F., et al., *Proc Natl Acad. Sci. U.S.A.* **90**: 11478-11482 (1993)].

The invention described hereinafter provides an autogeneic immunological process for the production of antibodies to CETP and can provide long-term
15 lessening of transfer of cholesteryl esters from HDL particles in mammals whose blood contains CETP by utilization of a DNA vaccine. This process permits the long-term elevation of anti-atherogenic HDL cholesterol concentrations.

20

Brief Summary of the Invention

The present invention contemplates an autogeneic immunological process for lessening the
25 transfer of cholesteryl esters from HDL particles and for increasing the HDL cholesterol concentration of a mammal whose blood also contains CETP. A contemplated process is useful in treating human pro-atherogenic dyslipoproteinemias characterized by low HDL/LDL
30 cholesterol ratios. Also contemplated here are isolated and purified DNA that encode a useful immunogen and expression systems for that DNA.

One contemplated process comprises the steps of:

(a) immunizing the mammal to be treated with an inoculum containing a DNA molecule that encodes a CETP immunogen that is an immunogenic polypeptide having a CETP amino acid residue sequence and which DNA molecule is dissolved or dispersed in a vehicle; and

(b) maintaining the immunized mammal for a time period sufficient for the immunizing DNA to express the immunogenic polypeptide to induce the production of antibodies that bind to CETP, and preferably also lessen the transfer of cholesteryl esters from HDL. In one embodiment, the DNA encodes an immunogenic polypeptide that is an intact CETP molecule such as recombinant human or rabbit CETP. In another embodiment, the encoded immunogenic polypeptide is a portion of a CETP molecule that is covalently bonded to an exogenous antigenic carrier as a fusion protein.

In preferred embodiments, the exogenous antigenic carrier is the hepatitis B core protein (HBcAg) or diphtheria toxoid. HBcAg is particularly preferred as an encoded exogenous antigenic carrier, that forms a fusion protein with an immunogenic polypeptide having an amino acid residue sequence of the carboxy-terminal 30 residues of CETP. That more preferred fusion protein constitutes a polypeptide having the amino acid residue sequence of the hepatitis B core antigen from which about 3 to about 53 amino acid residues have been deleted and replaced by the immunogenic polypeptide that more preferably still has a length about equal to the number of amino acid residues deleted from HBcAg. The resulting fusion protein is most preferably expressed as particles having the size of HBcAg particles (about 27 nm).

The present invention has several benefits and advantages. One salient benefit is that a contemplated process can be utilized to lessen the CE transfer from

HDL to LDL or VLDL, thereby increasing the concentration of anti-atherogenic HDL cholesterol.

5 An advantage of the invention is that a contemplated process can have an effect that lasts for months as compared to the short-term effects of the small molecule drugs now available.

10 Another benefit of a contemplated process is that it utilizes the host mammal's own (autogeneic) immunological system to provide a desired result, thereby obviating problems associated with repeated administration of xenogeneic antibodies that themselves become immunogenic in the host mammal.

15 Another advantage of some contemplated processes is that their use of well known and accepted exogenous antigenic carriers such as HBcAg, tetanus toxoid, and diphtheria toxoid can boost the host mammal's immunity to those pathogens.

20 Still further benefits and advantages of the present invention will become apparent to a skilled worker from the disclosure that follows.

Definitions

25 The term "recombinant" is used to denote version of a DNA, RNA, or protein molecule altered with respect to the native molecule and resulting from the deletion, substitution, or insertion into the chain, by chemical, enzymatic, or biological means, of a sequence (a whole or partial chain of DNA, RNA, or protein) not originally present in that chain.

30 The term "recombinant DNA molecule" is used to mean a hybrid DNA sequence comprising at least two nucleotide sequences not normally found together in nature.

35 The term "polypeptide" is used herein to denote a sequence of about 10 to about 500 peptide-

bonded amino acid residues. A whole protein as well as a portion of a protein having the stated minimal length are polypeptides.

5 The term "fusion protein" is used to denote the expression product of two or more different genes in which the amino acid residue sequences of both genes are expressed peptide-bonded together as a single molecule. It is noted that a fusion protein need not have the full length amino acid residue sequence of any protein, but rather usually contains two or more truncated sequences. 10 The term is therefore somewhat of a misnomer, but is nonetheless well known and used as defined here by those skilled in the art.

15 The term "fused", when referring to expression of a fusion protein, is used herein to mean peptide-bonded.

20 The term "whole length CETP" is used to denote the full length CETP molecule (for example 476 amino acid residues long for human CETP or 496 residues long for rabbit CETP) as available in nature or produced as a recombinant protein.

25 The term "CETP immunogen" is used to denote molecule that is used to induce the production of antibodies that immunoreact with (bind to) CETP.

30 The terms "immunogenic polypeptide having a CETP amino acid residue sequence" or "immunogenic polypeptide" are used to denote the anti-CETP antibody-inducing portion of a "CETP immunogen"; i.e., that portion of a CETP immunogen to which induced antibodies bind.

35 The term "exogenous antigenic carrier" or "carrier" is used herein to denote a molecule foreign to the immunized mammal that provides a signal to antibody-producing B cells. Such carriers and their functions are well known in the art. Such a carrier can be a

polypeptide having a sequence of as few as about 10 amino acid residues to the length of an intact protein, as well as being a synthetic polymer or oligomer.

5 The term "inoculum" in its various grammatical forms is used herein to describe a composition containing an amount of CETP immunogen (e.g., DNA encoding a polypeptide conjugate, CETP protein or recombinant protein) sufficient for a described purpose that is dissolved or dispersed in an aqueous,
10 physiologically tolerable diluent.

The term "expression" is used to mean the combination of intracellular processes, including transcription and translation undergone by a structural gene to produce a polypeptide.

15 The terms "operatively linked" or "operably inserted" are used to mean that two or more DNA sequences are covalently bonded together in correct reading frame.

The term "promoter" is used to mean a
20 recognition site on a DNA sequence or group of DNA sequences that provide an expression control element for a gene and to which RNA polymerase specifically binds and initiates RNA synthesis (transcription) of that gene.

25 The term "structural gene" is used to mean a DNA sequence that is expressed as a polypeptide; i.e., an amino acid residue sequence.

The term "vector" is used to mean a DNA molecule capable of replication in a cell and/or to
30 which another DNA segment can be operatively linked so as to bring about replication of the attached segment. A plasmid is an exemplary vector.

The term "expression vector" is used to mean a
35 DNA sequence that causes a polypeptide to be expressed in that the DNA sequence contains control elements that

regulate expression of structural genes when operatively linked to those genes within a vector.

Detailed Description of the Invention

5 The present invention relates to a process for
producing antibodies to CETP. Preferably, the produced
antibodies lessen the transfer of cholesteryl esters
from HDL, and increase the ratio of HDL cholesterol to
10 LDL cholesterol in the blood of a treated mammal that
has CETP in its blood. In humans, that increase in HDL
to LDL ratio can lead to an amelioration of
dyslipoproteinemias characterized by low HDL/LDL
cholesterol ratios. That desired raising of the HDL/LDL
15 cholesterol ratio is accomplished immunologically by
antibodies induced in the blood of the treated mammal
that recognize and bind to circulating CETP. Also
contemplated in this invention are a DNA that encodes an
immunogen utilized in the process, an inoculum that
utilizes the DNA, and an isolated and purified DNA
20 segment that encodes a contemplated immunogen.

I. The Process

 A contemplated process is referred to herein
as utilizing "autogeneic" antibodies to denote that the
25 useful antibodies are those induced in the host mammal
itself. This autogeneic immunological process is
therefore to be distinguished from a xenogeneic process
in which antibodies from an animal of one species are
administered to an animal of another species as where
30 mouse anti-CETP TP2 or 1C4 monoclonal antibodies have
been administered to hamsters or rabbits. A
contemplated autogeneic immunological process is also to
be distinguished from an allogeneic immunological
process such as a passive immunization in which
35 antibodies from one animal are administered to another

animal of the same species as where humans receive gamma globulin injections from other humans.

A contemplated process is thus closely analogous to an autoimmune process in which a mammal's own immune system attacks an endogenous or self protein. CETP is an endogenous protein in rabbits, hamsters and primates that are among the mammalian hosts contemplated here. However, inasmuch as the cause of most if not all autoimmune responses is presently unknown and the desired immune response contemplated here is purposefully induced, it is believed appropriate to use a different name for the result obtained here.

One contemplated process produces antibodies to CETP in a mammal. That process comprises the steps of:

(a) immunizing the mammal with an inoculum containing a vehicle in which is dissolved or dispersed a recombinant DNA molecule comprising a DNA sequence that contains (i) a sequence encoding a CETP immunogen linked to (ii) a promoter sequence that controls the expression of the CETP immunogen DNA sequence in the mammal. The encoded CETP immunogen is an immunogenic polypeptide having a CETP amino acid residue sequence. The immunization provides an amount of the recombinant DNA molecule sufficient to be expressed and for the expressed immunogenic polypeptide to induce antibodies to CETP. The immunized mammal is (b) maintained for a time period sufficient for production of antibodies that bind to CETP.

Another aspect of the present invention contemplates a process for lessening the transfer of cholesteryl esters from HDL particles and increasing the concentration of HDL cholesterol in the blood of a mammal whose blood contains cholesteryl ester transfer protein (CETP); i.e., animals that have endogenous

plasma CETP measured as a transfer activity. That process comprises the steps of: (a) immunizing that mammal (the host) with an inoculum that contains a DNA-encoded CETP immunogen and linked, controlling promoter sequence dissolved or dispersed in a vehicle. The DNA-encoded CETP immunogen is an immunogenic polypeptide having a CETP amino acid residue sequence. The immunized mammal is (b) maintained for a time period sufficient for the DNA to express the immunogenic polypeptide and for the expressed immunogenic polypeptide to induce the production of antibodies that bind to CETP, and preferably lessen the transfer of cholesteryl esters (CE) from HDL.

A. The CETP-Encoded Immunogen

The immunizing DNA that encodes an immunogenic polypeptide having a CETP amino acid residue sequence of the CETP immunogen can encode a whole CETP molecule such as the human (476 residues) or rabbit (496 residues) proteins whose amino acid residue sequences are provided as SEQ ID NOs:28 and 26, respectively, and whose DNA sequences are provided in SEQ ID NOs:1 and 27, respectively. The cDNA (SEQ ID NO:31) and deduced amino acid residue sequence (SEQ ID NO:30) for cynomolgus monkey CETP have also been reported by Pape et al., *Atherosclerosis and Thrombosis*, 11:1759-1771 (1991). A DNA that encodes a polypeptide of SEQ ID NO:30 or a portion thereof as described below, can also be utilized herein; i.e., the cDNA shown in SEQ ID NO:31 or a portion thereof, respectively.

Where the whole CETP molecule is encoded by the DNA as the immunogenic polypeptide of the CETP immunogen, it is preferred to use the DNA sequence that encodes a CETP sequence from an animal species other than that of the immunized mammal; i.e., the encoded

CETP is preferably xenogeneic as to the immunized mammal. When an encoded immunogenic polypeptide is other than an intact CETP molecule, it is preferred to use a DNA that encodes a polypeptide having a length of about 10 to about 30 amino acid residues, and more preferably, a length of about 20 to 30 residues. In this instance, the immunogenic polypeptide is expressed covalently (peptide) bonded to an exogenous antigenic carrier to form the CETP immunogen. The encoded immunogenic polypeptide and antigenic carrier sequences are, of course, linked together in proper reading frame, as is an encoded immunogenic polypeptide linked to the promoter when used without an antigenic carrier.

Exogenous antigenic carrier polypeptide molecules are also well known in the art, as are the amino acid residue and nucleotide sequences of those molecules. Exemplary polypeptide carriers include but are not limited to tetanus toxoid, diphtheria toxoid, thyroglobulin and the hepatitis B core protein (HBcAg).

Thus, the cDNA encoding an exogenous antigenic carrier and that encoding an immunogenic CETP polypeptide are operatively linked to form a single isolated and purified DNA molecule that encodes both the carrier and immunogenic polypeptide. That DNA molecule can then be operatively linked in an appropriate expression vector along with a promoter that controls expression of those two polypeptides as a single fusion protein whose two polypeptide portions are covalently bonded by a peptide bond. Preferably, the carrier is expressed at the amino-terminus of the fusion protein, although a carrier can also be expressed fused at the carboxy-terminus of the immunogenic polypeptide. Exemplary proteins and procedures for their synthesis are discussed hereinafter.

Preferably, where the whole CETP molecule is used as the immunogenic polypeptide, the carrier polypeptide has an amino acid residue sequence that is less than that of a whole protein. That length is preferably about 15 to about 70 amino acid residues.

The hepatitis B nucleocapsid or core protein antigen also referred to as HBcAg is a particularly preferred exogenous antigenic carrier, as will be discussed in greater detail hereinafter. The HBcAg molecule is often used illustratively herein as a carrier.

U.S. Patent No. 4,818,527, whose disclosures are incorporated by reference, teaches that the region extending from about position 70 through about position 140 from the amino-terminus of HBcAg, whose complete amino acid and cDNA sequences are shown as SEQ ID NOS:38 and 39, respectively, is particularly useful as a T cell independent stimulant as are sequences of about 15 to about 25 residues from that region. The amino acid residue sequences of four of those shorter polypeptides are provided as SEQ ID NOS:40, 41, 42 and 43. The cDNA sequences that encode each of those four polypeptides can be readily obtained from SEQ ID NO:39, and the 3' end of such a cDNA can be operatively linked to the 5' end of cDNA that encodes an immunogenic polypeptide, or vice versa, for expression as a fusion protein CETP immunogen.

Thus, in one embodiment, a preferred recombinant DNA molecule (defined previously and discussed hereinafter) encodes a CETP immunogen that is a fusion protein whose amino-terminal portion is a polypeptide having a length of about 15 to about 70 amino acid residues and having the sequence of HBcAg from about position 70 to about position 140 from the HBcAg amino-terminus. The carboxy-terminal portion of

that fusion protein has the amino acid residue sequence of a CETP molecule, and the two portions are covalently bonded by a peptide bond. In this embodiment, the CETP molecule can be from the same species as the immunized mammal.

In another preferred embodiment, the immunizing DNA encodes a CETP immunogen that is comprised of an exogenous antigenic carrier to which one or more immunogenic polypeptides having a length of about 10 to about 30 amino acid residues such as those of SEQ ID NOS:2-7 or 50 having a sequence of rabbit CETP, the similar polypeptides of SEQ ID NOS:8-13 or 29 having a sequence of human CETP or the similar polypeptides of SEQ ID NOS:32-37 having a sequence of monkey CETP is covalently bonded. Here, the carrier is preferably an intact protein such as a before-noted tetanus toxoid, diphtheria toxoid, thyroglobulin or HBcAg molecule.

As noted before, a DNA sequence for the CETP molecule or a desired portion thereof can be obtained as described by M.E. Pape et al., *Arteriosclerosis and Thrombosis*, 11:1759-1771 (1991); N.W. Jeong et al., *Mol. Cells*, 4(4):529-533 (1994); and D.T. Connolly et al., *Biochem. J.*, 320:39-47 (1996). Oligonucleotides can also be prepared using standard synthetic technology where shorter DNA sequences are desired. Those oligonucleotides can also be linked enzymatically, as with T4 DNA ligase, to form longer molecules.

DNA sequences for exogenous antigenic carrier molecules have also been reported as have methods for expressing those molecules. For example, a DNA sequence that encodes the preferred HBcAg exogenous antigenic carrier is disclosed in U.S. Patent No. 4,710,463, whose disclosures are incorporated herein by reference, and

E. coli-containing plasmids whose DNA encode hepatitis B virus proteins were deposited in the Culture Collection of the National Collection of Industrial Bacteria, Aberdeen Scotland as pBR322-HBV G-L. In addition, DNA
5 encoding HBcAg is disclosed in U.S. Patent No. 4,942,125 as present in vectors deposited at the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, MD 20852-1776 as ATCC No. 39629, No. 39631 and No. 40102.

10 The use of HBcAg as an exogenous antigenic carrier in a fusion protein is illustrated in Moriarty et al., *Vaccines 90*, Brown et al. eds., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 225-229 (1990). The authors there reported operatively
15 linking the 3' end of DNA encoding a 17-mer amino acid residue sequence of the HIV *gag* protein to the 5' end of DNA encoding HBcAg, and reported that appropriately transfected *E. coli* expressed a fusion protein having the HIV *gag* sequence peptide-bonded to the amino-
20 terminus of HBcAg. That expressed fusion protein was present in particulate form and was shown to be an excellent immunogen in mice.

 Schödel et al., *Vaccines 90*, Brown et al. eds., Cold Spring Harbor Laboratory Press, 193-198 Cold
25 Spring Harbor, N.Y. (1990) reported the preparation and successful use of a fusion protein immunogen that contained a polypeptide immunogen having an amino acid residue sequence of hepatitis B Pre-S2 (residues 133-140) that was expressed peptide-bonded to the carboxy-
30 terminus of HBcAg so that the 3' end of the exogenous carrier (HBcAg) DNA was linked to the 5' end of the DNA that encoded the Pre-S2 polypeptide immunogen. That expressed fusion protein immunogen was also obtained in particulate form.

Similar techniques can be utilized here using a CETP-encoding recombinant DNA molecule that contains a DNA molecule of SEQ ID NOS:14-19, 20-25 or a corresponding DNA sequence of SEQ ID NO:31 that encodes a CETP immunogenic polypeptide in place of the DNAs used by the Moriarty et al. or the Schödel et al. groups.

In addition, using similar techniques and others well known to workers of ordinary skill in the recombinant DNA art, a DNA molecule that encodes a fusion protein can be prepared that expresses a polypeptide having an HBcAg amino acid residue sequence such as one of those of SEQ ID NOS:40-43 peptide-bonded to the amino-terminus of an intact CETP molecule.

A particularly preferred DNA-encoded CETP immunogen is a fusion protein comprised of an immunogenic polypeptide having a length of 10 to about 30 amino acid residues that is peptide-bonded to both an amino-terminal flanking amino acid residue sequence and a carboxy-terminal flanking sequence, and is sometimes referred to hereinafter HBcAg/CETP/HBcAg. Those flanking sequences are preferably portions from the amino-terminal and carboxy-terminal regions of the HBcAg molecule, as was discussed previously. Thus, in this fusion protein, the exogenous antigenic carrier molecule is encoded to be peptide-bonded to both the amino-terminus and carboxy-terminus of the immunogenic polypeptide.

A preferred encoded polypeptide immunogen (immunogenic polypeptide) has an amino acid residue sequence of SEQ ID NOS:2-7, 8-13, 29, 32-37 or 50. Most preferably, the encoded polypeptide immunogen has an amino acid residue sequence that is bound by (immunoreacts with) the monoclonal antibodies designated TP1, TP2 and TP3 reported by B. Hessler et al., *J. Biol. Chem.*, 263(11):5020-5023 (1988), or that denominated 1C4

by J. Gaynor et al., *Atherosclerosis*, 110(1):101-109
(1994). Monoclonal antibody TP2 binds to an epitope
located between about positions 465 and 475 of human
CETP. Tall, J. *Lipid Res.*, 34:1255-1274 (1993), and the
5 citations therein.

A particularly preferred recombinant DNA
molecule contains a controlling promoter sequence linked
to an encoded polypeptide immunogen sequence whose
encoded amino acid residue sequence includes positions
10 465 through 475 of human CETP or an analogous position
of CETP from another source. An encoded polypeptide is
exemplified by those of SEQ ID NOs:4, 10, 29 and 34, of
which the encoded polypeptides of SEQ ID NOs:10 and 29
are most preferred. The polypeptide of SEQ ID NO:10 is
15 encoded by the DNA of SEQ ID NO:22.

Protein molecules have not only a linear amino
acid residue or primary sequence, but also can possess a
secondary sequence in which the polypeptide back bone is
coiled in an α -helix or folded into a β -sheet, as well
20 as a tertiary sequence in which sequentially distant
portions of the molecule are folded to be adjacent to
each other. Many linear antigenic/immunogenic
polypeptide sequences have been reported in the
literature, and such sequences can be readily mimicked
25 by polypeptides having a length of 10 to about 30 amino
acid residues. Such relatively short polypeptides
typically do not mimic a secondary structure such as an
 α -helix in aqueous media.

The region of CETP that immunoreacts with
30 monoclonal antibody TP2 is predicted to have an
amphipathic helical secondary structure, with the
hydrophilic surface bound by the antibody. See Wang et
al., *J. Biol. Chem.*, 267(25):17487-17490 (1992) and A.R.
Tall, J. *Lipid Res.* 34:1255-1274 (1993). A contemplated

DNA encodes a CETP immunogen fusion protein having an immunogenic polypeptide flanked at its amino- and carboxy-termini by peptide-bonded regions of HBcAg; i.e., HBcAg/CETP/HBcAg, that is more constrained in its molecular motions than is an immunogenic polypeptide that is bonded at only one terminus. As a consequence, by flanking a before-mentioned particularly preferred immunogenic polypeptide with regions of HBcAg to form a HBcAg/CETP/HBcAg fusion protein, it is believed that the expressed immunogenic polypeptide becomes constrained to have a helical structure much like that present in the native CETP molecule and thereby induce autogeneic antibodies having an antigenic specificity similar to those exhibited by mouse monoclonal antibodies TP1, TP2, TP3 and 1C4 discussed previously.

It is further believed that formation of HBcAg-like particles of an expressed fusion protein HBcAg/CETP/HBcAg immunogen places further conformational constraints upon the immunogenic polypeptide by which the immunogenic polypeptide becomes the primary immunogen with loss of much of the HBcAg immunogenicity, while the T cell-independent antigenic carrier function of HBcAg is retained. See Schödel et al., *J. Virol.*, 66(1):106-114 (1992) for a similar result using a different immunogen.

Although use of the full length HBcAg exogenous antigenic carrier molecule or substantially full length molecule has thus far been discussed, it is noted that about 10 amino-terminal amino acid residues (about 30 base pairs) or about 40 carboxy-terminal amino acid residues (about 120 base pairs) can be deleted from the expressed HBcAg/CETP/HBcAg sequence (encoding DNA) without abrogating function as an exogenous antigenic carrier or assembly into particles. See, for example, Birnbaum et al., *J. Virol.*, 64(7):3319-3330 (1990).

Exemplary preparations of immunogenic fusion proteins having HBcAg as a carrier with various heterologous polypeptide insertions from pathogens as immunogen, and also usage of full length and carboxy-terminal deletions in the HBcAg amino acid residue sequence can be found in the following publications. Schödel et al., *J. Exp. Med.*, 180:1037-1046 (1994); Schödel et al., *J. Virol.*, 66(1):106-114 (1992); Schödel et al., *Vaccines 91*, Brown et al. eds., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 319-325 (1991); Clarke et al., *Vaccines 91*, Brown et al. eds., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 313-318 (1991); and Schödel et al., *Vaccines 90*, Brown et al. eds., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 193-198 (1990).

It is also noted that the human hepatitis virus (HBV), whose core antigen is discussed herein, has two subtypes that are denominated adw and ayw. The core antigens of those two viral subtypes have slightly different DNA and amino acid residue sequences. Although subtype specificity has been noted as to the immunogenicity of the S and PreS regions of HBV, [see, for example, Milich et al. *Vaccines 86*, Brown et al. eds., Cold Spring Harbor Press, Cold Spring Harbor, N.Y., 377-382 (1986)] either subtype can be used as an exogenous antigenic carrier herein, with subtype ayw being used illustratively herein.

It should also be noted that although a contemplated process has to a great extent been discussed in terms of the polypeptide immunogen that is ultimately expressed and induces autogeneic antibody production to CETP, that immunogenic polypeptide may not be readily discernable in an immunized host mammal. The

DNA that encodes such an immunogenic polypeptide may also not be easily identifiable in the mammalian host where no reporter gene such as β -galactosidase is included as a part of the immunizing recombinant DNA molecule, or where an immunizing recombinant DNA molecule, which exists and functions extrachromosomally, is eliminated from an immunized cell at a time subsequent to expression of a CETP immunogen. Antibodies that bind to CETP are, however, present in the host after an appropriate amount of time, and the presence of those antibodies provides evidence that a contemplated process has been carried out in that such CETP-binding antibodies are not known to arise naturally.

15 B. DNA Molecules and Expression Systems

 A contemplated DNA molecule (isolated purified DNA segment) that encodes a CETP immunogen can be referred to as a number of base pairs at a particular location in a plasmid, as a restriction fragment bounded by two restriction endonuclease sites, and as a restriction fragment bounded by two restriction endonuclease sites and containing a number of base pairs. A contemplated DNA can also be defined to have a sequence of a denominated SEQ ID NO, as well as alleles or variants of such genes (described hereinafter) that encode a recited amino acid residue sequence.

 A contemplated isolated and purified DNA segment is linear, and as such has a 5' end and a 3' end. A contemplated DNA segment can comprise two or more individual DNA segments whose 3' ends are operatively linked to the 5' end of another DNA segment where two segments are joined, or whose 3' end is operatively linked to the 5' end of another DNA segment whose own 3' end is operatively linked to the 5' end of

yet another DNA segment, where three individual segments are joined to form a single isolated and purified DNA segment.

5 In living organisms, the amino acid residue sequence of a protein or polypeptide is directly related via the genetic code to the deoxyribonucleic acid (DNA) sequence of the structural gene that codes for the protein. A structural gene can be defined in terms of the amino acid residue sequence; i.e., protein or
10 polypeptide, for which it codes.

In addition, through the well-known redundancy of the genetic code, additional DNA sequences can be prepared that encode the same amino acid residue sequences, but are different from a recited gene
15 sequence having a particular SEQ ID NO. For example, *in vitro* mutagenesis as is illustrated hereinafter can be used to change a DNA sequence so that the same residue of an expressed polypeptide is expressed using one or more different codons. In addition, that same technique
20 can be used to change one amino acid residue to another where it is desired to insert or delete specific restriction endonuclease sites. This technique is also illustrated hereinafter.

A DNA sequence that encodes a CETP immunogen
25 of a recited SEQ ID NO but has a DNA sequence different from that of a recited SEQ ID NO is referred to herein as a variant DNA sequence. Such a variant DNA molecule can be readily prepared by *in vitro* mutagenesis, as is well known.

30 A DNA segment that encodes a described CETP immunogen can be synthesized by chemical techniques, for example, the phosphotriester method of Matteucci et al., *J. Am. Chem. Soc.*, 103:3185 (1981). Of course, by chemically synthesizing the coding sequence, any desired

modifications can be made simply by substituting the appropriate bases for those encoding the native amino acid residue sequence.

5 However, DNA segments including the specific sequences discussed previously are particularly preferred. Furthermore, a DNA segment that encodes a polypeptide can be obtained from a recombinant DNA molecule (plasmid or other vectors) containing that segment.

10 A DNA segment that includes a DNA sequence encoding a CETP immunogen can be prepared by excising and operatively linking appropriate restriction fragments from appropriate plasmids or other DNA using well known methods. The DNA molecules useful here that
15 are produced in this manner typically have cohesive termini; i.e., "overhanging" single-stranded portions that extend beyond the double-stranded portion of the molecule. The presence of cohesive termini on the DNA molecules of the present invention is preferred,
20 although molecules having blunt termini are also contemplated.

 A recombinant DNA molecule useful herein can be produced by operatively linking a vector to an isolated DNA segment that encodes a CETP immunogen to
25 form a plasmid such as those discussed herein. Particularly preferred recombinant DNA molecules are discussed in detail in the examples, hereafter. Vectors capable of directing the expression of the gene in the immunized mammal are referred to herein as "expression
30 vectors".

 The expression vectors described above contain expression control elements including a promoter. The genes that encode an immunogenic polypeptide or other useful sequence are operatively linked to the expression
35 vector to permit the promoter sequence to direct RNA

polymerase binding and expression of the desired polypeptide coding gene.

5 The choice of which expression vector to which a polypeptide-coding gene is operatively linked depends directly on the functional properties desired, e.g. the location and timing of protein expression, and the host cell to be transformed. These are well known limitations inherent in the art of constructing recombinant DNA molecules. However, a vector useful in practicing the present invention is capable of directing the replication and also the expression of the immunogenic polypeptide-coding gene included in the DNA segment to which it is operatively linked.

10 A cloning vector is also useful herein for making or increasing the amount of a desired DNA, and can also be used to express an immunogenic polypeptide to assay the synthesized DNA. A cloning vector includes a prokaryotic replicon; i.e., a DNA sequence having the ability to direct autonomous replication and maintenance of the recombinant DNA molecule extrachromosomally in a prokaryotic host cell transformed therewith. Such replicons are well known in the art as are cloning vectors, some of which are discussed below.

15 Those vectors that include a prokaryotic replicon can also include a prokaryotic promoter region capable of directing the expression of the CETP immunogen gene in a host cell, such as *E. coli*, transformed therewith. Promoter sequences compatible with bacterial hosts are typically provided in plasmid vectors containing one or more convenient restriction sites for insertion of a DNA segment of the present invention. Such cloning and expression vector plasmids are well known in the art. Typical of such cloning and expression vector plasmids are pUC18, pUC19, pBR322, pProEx1, and pFastBac1 available from Life Technologies,

Rockville, MD, and pPL and pKK223-3 available from Pharmacia, Piscataway, N.J. These vectors are utilized in the synthesis of the DNA segments useful herein.

5 In preferred embodiments, the cloning vector used to express an immunogenic polypeptide-coding gene includes a selection marker that is effective in a host cell, preferably a drug resistance selection marker. One preferred drug resistance marker is the gene whose expression results in kanamycin resistance, whereas
10 ampicillin resistance is another such marker. Again, such selective markers are well known.

A variety of methods has been developed to operatively link DNA to vectors via complementary cohesive termini or blunt ends. For instance,
15 complementary homopolymer tracts can be added to the DNA segment to be inserted and to the vector DNA. The vector and DNA segment are then joined by hydrogen bonding between the complementary homopolymeric tails to form recombinant DNA molecules.

20 Alternatively, synthetic linkers or adapters containing one or more restriction endonuclease sites can be used to join the DNA segment to the integrating expression vector. The synthetic linkers or adapters are attached to blunt-ended DNA segments by incubating
25 the blunt-ended DNA segments with a large excess of synthetic linker or adapter molecules in the presence of an enzyme that is able to catalyze the ligation of blunt-ended DNA molecules such as bacteriophage T4 DNA ligase.

30 Thus, the products of the reaction are DNA segments carrying synthetic linker sequences at their ends. These DNA segments are then cleaved with the appropriate restriction endonuclease and ligated into an expression vector that has been cleaved with an enzyme
35 that produces termini compatible with those of the

synthetic linker. Synthetic linkers containing a variety of restriction endonuclease sites are commercially available from a number of sources including New England BioLabs, Beverly, MA. A synthetic adapter molecule typically has sticky end and one blunt end and is not cleaved after ligation.

Although preferred, it is not always feasible to design a DNA molecule whose expressed polypeptide has the exact terminal residues of a polypeptide enumerated in a SEQ ID NO. This is because of the limitations inherent in the use of restriction enzymes, synthetic linkers and adapter molecules used for cutting and joining DNA segments.

As a consequence, an expressed polypeptide can contain a few (e.g., one or two) more, less or different amino acid residues at one or both termini of an enumerated sequence. Such slight changes are well tolerated by a contemplated CETP immunogen, particularly when the substitution is conservative and residues such as Cys and Pro are avoided.

A variety of plasmids can be used as DNA vaccine vectors for expressing a contemplated CETP immunogen in a mammalian host. Such vectors optimally include the following components: a strong eukaryotic promoter, a cloning site for insertion of a gene of interest, a polyadenylation termination [poly(A)] sequence, a prokaryotic origin of replication, and a prokaryotic selectable marker. One such vector, pV1J, contains the cytomegalovirus immediate-early promoter with intron A, a bovine growth hormone polyadenylation termination sequence, and an ampicillin resistance gene. [J.B. Ulmer et al., *ASM News*, 62(9): 476-479 (1996).] Another useful vector denominated pcDNA1/Amp that is available from Invitrogen, Corp. of San Diego, CA, as well as plasmids pCMV-SPORT- β -gal and pGreen Lantern-1

available from Life Technologies, Rockville, MD are discussed in detail hereinafter. Other eukaryotic promoters, poly(A) sites, and selectable markers can be substituted without departing from the utility of the vector, as long as the structural gene inserted downstream from the promoter is expressed in mammalian cells. A variety of general mammalian expression vectors, many of which are commercially available, are suitable for use herein as DNA vaccine vectors.

Plasmid DNA can be prepared by a variety of methods. High quality plasmid DNA can be prepared using CsCl gradients to separate covalently closed circular plasmid DNA from linearized plasmid and chromosomal DNA. Other components from lysed bacterial cells such as proteins, RNA, membranes, and cell wall material are generally well-separated from the plasmid DNA by density gradient centrifugation. Standard protocols for CsCl purification are well known in the art. [See, J. Sambrook et al., Molecular Cloning, 2nd ed., Cold Spring Harbor Press, Cold Spring Harbor N.Y., 8-23 (1989).]

Another method suitable for preparation of high quality plasmid DNA is anion exchange chromatography. Direct comparisons of DNA prepared by anion exchange chromatography and DNA banded twice on CsCl gradients reportedly showed no difference in the efficiency of direct gene transfer and genetic immunization for reporter plasmids. [H.L. Davis et al., *Biotechniques*, 21: 99-99 (1996).] DNA banded once on CsCl gradients was reported to contain less double-stranded closed circular DNA and more RNA contamination than DNA banded twice on these gradients. Endotoxin levels were said to be greater, however, for DNA purified by anion exchange chromatography than by CsCl purification. [H.L. Davis et al., *Biotechniques*, 21: 99-99 (1996).]

A preparative method of purification of supercoiled plasmid DNA for therapeutic applications can also be used [A.P. Green et al., *Biopharm*, 10(5): 52-62, (1997)]. This method uses ion-pairing reversed phase column chromatography to prepare plasmid DNA free from contaminating DNA, RNA, protein, and endotoxin.

General methods and procedures for characterizing bulk plasmid DNA vectors suitable for use in human gene therapy have been described [M. Marquet et al., *BioPharm*, 10(5): 42-50 (1997)]. Methods to characterize a master cell bank, by testing plasmid identity, plasmid yield, plasmid stability, and methods to verify the genotype of the host strain and to measure host strain viability and microbial contamination were described. Methods to characterize the bulk product in terms of sterility and for the presence of pyrogens were also described.

Endotoxin is the lipopolysaccharide component of the cell wall of gram-negative bacteria that is released when cells are lysed for recovery of plasmid DNA. Endotoxin can have cytotoxic effects on mammalian cells in vitro or in vivo. [I.P. Wicks et al., *Hum. Gene Ther.*, 6: 317-323 (1995)]. Endotoxin levels can be measured by a chromogenic limulus amoebocyte clotting assay [SCL-100 kit; BioWhittaker, Walkerville, MD; T. Mossman, *J. Immunol. Meth.*, 65: 55-63 (1983)]. Methods for purification of endotoxin-free plasmid DNA prepared by anion exchange chromatography are now available. [J. Schorr et al., *Gene Ther.*, 1: S7 (1994)]. This method clears the bacterial lysate with a special endotoxin removal buffer before the DNA is purified from lysate on an anion exchange column.

A plasmid or other vector DNA that encodes a contemplated CETP immunogen and contains a promoter sequence as discussed before that controls expression of

that immunogen-encoding DNA sequence in proper reading
frame comprises a recombinant DNA molecule utilized in
immunizing a host mammal. That recombinant DNA molecule
dissolved or dispersed in a vehicle comprises an
5 inoculum that is used to immunize the mammal and is
discussed further hereinbelow.

C. Inocula

A DNA vaccine vector encoding a CETP immunogen
10 in proper reading frame (a recombinant DNA molecule) and
containing a promoter sequence that controls expression
of the immunogenic polypeptide is dissolved or dispersed
in a pharmaceutically-acceptable vehicle composition
that is preferably aqueous to form an inoculum that when
15 used to immunize a mammal induces the production of
antibodies that immunoreact with (bind to) CETP. When
that recombinant DNA molecule is administered in an
effective amount to a mammal whose blood contains CETP
those antibodies preferably also lessen the transfer of
20 cholesteryl esters from HDL particles.

An effective recombinant DNA molecule dosage
is typically about 0.05 μ g/kg to about 50 mg/kg, usually
about 0.005 mg/kg to about 5 mg/kg. Methods of
determining the effective systemic dose, which dose can
25 vary depending on the activity of the polypeptide
encoded by the DNA can be determined in a manner
apparent to those of skill in the art. U.S. Patent No.
5,580,859, whose disclosure is herein incorporated by
reference, discloses several methods of determining an
30 effective dose of a DNA vaccine.

The term "unit dose" as it pertains to an
inoculum of the present invention refers to physically
discrete units suitable as unitary dosages for animals,
each unit containing a predetermined quantity of active
35 material calculated to individually or collectively

produce the desired immunogenic effect in association with the required diluent; i.e., carrier, or vehicle.

Inocula are typically prepared from a recombinant DNA molecule encoding a CETP immunogen by dispersing the DNA in a liquid physiologically tolerable (acceptable) diluent vehicle such as water, or phosphate-buffered saline (PBS), or the like to form an aqueous composition. The amount of DNA-encoding CETP immunogen utilized in each immunization can vary widely, and is referred to as an effective amount. Such an effective amount is sufficient to induce antibodies to CETP that bind to CETP, and preferably lessen the transfer of cholesteryl esters from HDL particles, and also increase the HDL/LDL ratio in the immunized mammal's blood. Exemplary effective amounts of a DNA-encoded CETP immunogen are about 0.005 mg/kg to about 5 mg/kg, depending inter alia, upon the sequence of the encoded CETP immunogen, the mammal immunized, and the presence of salts, stabilizers, and cell penetration agents in the inoculum. An exemplary unit does can constitute about 10 μ l to about 1 ml per site of administration (immunization), with a recombinant DNA molecule concentration of about 0.05 μ g/ml to about 20 mg/ml, and preferably about 0.1 μ g/ml to about 100 μ g/ml. Thus, a single unit dose or a plurality of unit doses can be used to provide an effective amount of expressed CETP immunogen. Those skilled in the art know that appropriate concentrations or amounts can be readily determined.

An inoculum is typically formulated for parenteral administration. Exemplary immunizations are carried out sub-cutaneously (s.c.) intra-muscularly (i.m.) or intra-dermally (i.d.). U.S. Patent No. 5,580,859, discloses several formulations suitable for use with a DNA vaccine. These include inocula composed

of recombinant DNA molecules dissolved or dispersed in an aqueous vehicle containing salts, diluents, stabilizers, and cell penetration agents.

5 Typical formulations include recombinant DNA molecules suspended in phosphate buffered saline or isotonic sucrose. DNA can also be complexed with cell penetration agents such as liposomes, which facilitate uptake of the DNA into cells. One example of a liposome which has been used in DNA vaccine formulations is
10 Lipofectin™, available commercially from Life Technologies, Rockville, MD. Uptake and expression are often also significantly enhanced if DNA is administered in conjunction with the facilitating agent bupivacaine-HCl (an anesthetic) [Coney, L., et al., *Vaccine*, 12:
15 1545-1550 (1994)]. Those skilled in the art will recognize that other similar cell penetration and facilitating agents can be used for these purposes and can that appropriate concentrations or amounts of these agents can be readily determined.

20 Once immunized, the mammal is maintained for a period of time sufficient for the encoded CETP immunogen to be expressed and then for the expressed CETP immunogen to induce the production of antibodies that bind to CETP and lessen the transfer of cholesteryl
25 esters from HDL particles. This maintenance time typically lasts for a period of about three to about eight weeks, and can include a booster, second immunizing administration of the inoculum.

The production of antibodies that bind to CETP
30 is readily ascertained by obtaining a plasma or serum sample from the immunized mammal and assaying the antibodies therein for their ability to bind to CETP as an antigen in an ELISA assay as described hereinafter, or by another immunoassay such as a Western blot as is
35 well known in the art.

The lessening of transfer of cholesteryl esters from HDL can be assayed by one or more of several techniques. In one assay, the rate of transfer is measured by use of a [³H]-cholesteryl ester ([³H]CE) from HDL to LDL following the differential precipitation assay reported by Glenn et al., *Methods in Enzymology*, 263:339-350 (1996). Briefly, in a volume of 200 µl, CETP, [³H]CE-labeled HDL, LDL, and TES assay buffer (50 mM Tris, pH 7.4; 150 mM NaCl; 2 mM EDTA; 1% bovine serum albumin) are incubated for 2 hours at 37°C in 96-well filter plates. LDL is then differentially precipitated by the addition of 50 µl of 1% (w/v) dextran sulfate/0.5 M MgCl₂. After filtration, the radioactivity present in the precipitated LDL is measured by liquid scintillation counting. Correction for non-specific transfer or precipitation is made by including samples that did not contain CETP. The rate of [³H]CE transfer is determined in the linear range of the assay with respect to time and CETP concentration. For studies in which antibodies are included in the assay, the order of addition into sample wells is: buffer, [³H]CE-labeled HDL, LDL, antibodies, CETP.

CETP activity can also be measured using two methods that do not involve differential precipitation. In the first assay, the incubation conditions are identical to those described above, but separation of LDL acceptor particles from [³H]CE-labeled HDL donor particles is accomplished by size exclusion chromatography on tandem columns of Superose™ 6 (Sigma Chemical Co.), followed by liquid scintillation counting of fractions to determine the amount of [³H]CE associated with LDL and HDL. The amount of transfer measured by this method is typically in excellent agreement with the precipitation assay.

Another assay for CETP activity measures the rate of CETP-mediated transfer of the fluorescent analog NBD-cholesteryl linoleate (NBD-CE) from an egg phosphatidyl choline emulsion to VLDL. This assay takes advantage of the fact that NBD-CE is self-quenched when in the emulsion, and becomes fluorescent when transferred to VLDL. The assay is carried out according to the manufacturer's instructions (Diagnescent Technologies Inc., Yonkers, New York). Fluorescence measurements can be taken using a standard machine such as an SLM 8000C spectrophotofluorometer (Milton Roy Co., Rochester, New York) using 465 nm and 535 nm for excitation and emission wavelengths, respectively.

It is particularly contemplated once the desired antibodies are induced in the mammal that the immunization step be repeated at intervals of about 3 to about 6 months until the HDL cholesterol value in the blood of the mammal is increased by about 10 percent or more relative to the HDL cholesterol value for the mammal prior to the first immunization step. Preferably, the HDL cholesterol value is increased by about 25 percent. The mammal is thereafter preferably maintained at that increased HDL cholesterol level by periodic booster immunizations administered at intervals of about 6 to about 18 months. The increase in HDL cholesterol can be measured by any reliable assay, many of which are well known in the art, and one of which is described hereinafter.

It is noted that the before-described anti-CETP antibodies so induced can be isolated from the blood of the host mammal using well known techniques, and then reconstituted into a second inoculum for passive immunization as is also well known. Similar techniques are used for gamma-globulin immunizations of humans. For example, antiserum from one or a number of

immunized hosts can be precipitated in aqueous ammonium sulfate (typically at 40-50 percent of saturation), and the precipitated antibodies purified chromatographically as by use of affinity chromatography in which CETP or an immunogenic polypeptide portion thereof is utilized as the antigen immobilized on the chromatographic column.

Best Mode for Carrying Out The Invention

Comparative Examples 1 and 2, below, illustrate results obtained using a CETP polypeptide immunogen prepared exogenously to the immunized mammal.

Comparative Example 1: Immunization Of Rabbits With Rabbit CETP-Peptides

There is a 88 percent homology between rabbit and human CETP at the amino acid residue level. Rabbits express high levels of CETP in their blood and were chosen as a model for illustrating production of autogeneic anti-CETP antibodies.

The six rabbit CETP polypeptides of SEQ ID NO:2-7 were selected for this study and were prepared by standard solid phase synthesis procedures discussed below. To enhance the anti-polypeptide-specific antibody responses, two separate immunization strategies were used with the above six rabbit CETP-polypeptides.

A. Immunization Strategy 1 (MAP conjugates)

Rabbit polypeptides were synthesized as multiple antigenic peptide (MAP) constructs [D. N. Posnett et al., J. Biol Chem., 263:1719-1725 (1988)]. Those polypeptides were separately covalently bonded to "oligolysine core" molecules that were themselves covalently attached to resin particles [S. Butz et al., Pep. Res. 1:20-223 (1994)].

The substitution of the starting resin particles was 0.37 μm sites/mg resin that provided approximately 500 μg of immunogenic polypeptide per 1.1 mg resin. For the preparation of the CETP immunogen for immunization, 3.0 mg of dry resin were weighed out and hydrated in 1.3 ml sterile phosphate-buffered saline (PBS; pH 7.4) to which 1.3 ml Freund's complete adjuvant (CFA; Sigma Chemical Co., St. Louis, MO., F-5881) were added as adjuvant. The CETP immunogen and adjuvant were emulsified by a female-female luer lock syringe adapter connected to two 3 ml syringes. Each final emulsion was divided into 1.0 ml aliquots for injection (1 ml/rabbit), with one immunogen used per rabbit. Pre-immune rabbit serum was collected before immunization and stored at -70°C until immunoassay. On day 1, New Zealand white rabbits were separately immunized with respective immunogens by sub-cutaneous (s.c.) route on the back of the rabbit using 10 injection sites.

Three weeks later (on Day 22), the rabbits were boosted using similar procedures, but this time CETP immunogens were emulsified in Freund's incomplete adjuvant (IFA; Sigma). The resin-bonded CETP immunogen was weighed out as before and hydrated with sterile PBS the day before the booster immunization. The resulting CETP immunogen suspension was sonicated with a microtip at maximum setting for 5 minutes and left overnight (about 18 hours) at 4°C . Before mixing the hydrated CETP immunogen suspension with IFA, the suspension was warmed to room temperature just before the booster immunization, added to 1.5 ml IFA, and emulsified as described above to form an inoculum in which the CETP immunogen was dispersed. Rabbits were immunized each

with 1 ml of emulsion in at least 10 injection sites
s.c.

The first post-immune serum was collected 2
weeks after the second immunization from each animal.

5 All the anti-sera samples were stored in -70°C until
ELISA was done.

Using this MAP strategy, polypeptides of SEQ
ID NOS:2 and 7 were moderately immunogenic in rabbits
and resulted in maximum autogeneic antibody titers of
10 1:1000 and 1:300, respectively. The titers represent
the dilution of the sera that gave a half maximal
absorbance on ELISA plates coated with the respective
polypeptides. Sera were pooled from two rabbits, and
the above titers represent the mean value. Only anti-
15 sera to SEQ ID NO:7 cross-reacted with recombinant human
CETP. The reactivity of these anti-sera with rabbit
CETP is under evaluation using various immunological
assays. Anti-polypeptide-specific IgG has been purified
from the post-immune sera and its inhibitory property on
20 human recombinant CETP is being assayed.

B. Immunization Strategy 2 (Purified
Protein Derivative Conjugates)

Five of the above six rabbit CETP-polypeptide
immunogens (SEQ ID NOS:2, 3, 4, 6 and 7) were coupled to
25 tuberculin purified protein derivative (PPD) according
to the teachings of P.J. Lachmann et al., in 1986
Synthetic Peptides As Antigens, (Ciba Foundation
Symposium 119), 25-40 (1986) and P. Dawson et al., J.
Bio. Chem., 264:16798-16803 (1989) to form a conjugate.
30 The tuberculin PPD (Statens Serum Inst., Copenhagen,
Denmark) was used as an exogenous antigenic carrier to
enhance the immunogenicity of rabbit CETP-derived
polypeptides. The polypeptide-PPD conjugate in PBS was
emulsified with CFA as described for immunization

strategy 1. One ml of 0.5 mg/ml polypeptide conjugated to PPD was emulsified with approximately 1 ml CFA. A second 1 ml PPD-conjugate was frozen for next booster immunization.

5 On Day 1, rabbits were immunized with 1 ml of final emulsion in at least 10 sites sub-cutaneously on back of the rabbit. The polypeptide-PPD CETP polypeptide immunogen dose contained 0.25 mg of polypeptide per rabbit. Three weeks later (on Day 21),
10 the rabbits were given the booster immunization dose with the remaining 1 ml conjugate thawed and emulsified with IFA, as discussed before. Two weeks following the second immunization rabbits were bled to collect post-immune sera.

15 The PPD conjugation strategy resulted in antibodies to the immunogenic polypeptides of SEQ ID NOs:2 and 6, with antibody titers of 1:3200 and 1:400 respectively. The titers represent the dilution of the sera that gave a half maximal absorbance on peptide
20 coated ELISA plates. Sera were pooled from two rabbits and represent the mean value. Only the antibodies to the immunogenic polypeptide of SEQ ID NO:2 cross-reacted with recombinant human CETP. These results were
25 unexpectedly good inasmuch as P.J. Lachmann et al., supra, obtained substantially no anti-polypeptide antibodies in BCG-naïve hosts as were these rabbits. Anti-PPD antibodies were detected in all groups of rabbits as expected.

30 Using ELISA, the anti-immunogenic polypeptide sera are being used to evaluate their immuno-reactivity with natural rabbit CETP. Because the polypeptides of SEQ ID NOs:2, 6 and 7 were immunogenic and the two
35 anti-polypeptide antibodies against SEQ ID NOs:2 and 7 immunologically cross-reacted with recombinant human CETP, the respective rabbits were further boosted with a

third immunization dose either with the MAP constructs or PPD constructs emulsified with IFA.

Example 2: Immunization Of Outbred Rabbits
5 With CETP-Based Antigen

 This study utilized 30 New Zealand white rabbits in three groups with 10 rabbits per group. Three immunogens were utilized in this study: (1) Recombinant human CETP, (2) the carboxy-terminal 26
10 amino acid residues of rabbit CETP (SEQ ID NO:50), and (3) a control immunogen whose amino acid residue sequence was unrelated to that of CETP.

 Pre-immune sera were collected before immunization with the respective immunogens. The
15 purpose of this study was to illustrate that the above CETP immunogens would induce anti-CETP-specific (autogeneic anti-CETP) antibodies in rabbits, and that the autogeneic antibodies generated against CETP bind to (immunoreact with) the endogenous rabbit CETP, and thus
20 lessen the transfer of cholesteryl esters from HDL particles and raise the level of HDL in the hosts.

 The above immunogens were emulsified in CFA. Each rabbit received 500 µg of one of the immunogens emulsified in CFA immunized by sub-cutaneous route.
25 Seven weeks later the first bleed post-immune sera were collected.

 ELISA was employed to titrate the antibodies. ELISA plates were coated (40 ng/well) with the recombinant human CETP.

30 The rabbits immunized with recombinant human CETP exhibited a primary immune response against human CETP. All the ten rabbits responded well to the recombinant human CETP (rhCETP). The specific IgG antibody titer was >1:1000. However, the group of 10

rabbits immunized with the rabbit CETP carboxy-terminal polypeptide-thyroglobulin conjugate (CETP-TH) did not exhibit a primary antibody response. The control rabbit sera had no detectable levels of anti-CETP antibodies.

5 The rabbits were boosted with each respective antigen to further study immunogenicity.

The results of this study on the elevation of HDL particle concentration in the blood (plasma) of the host mammals (mean \pm S.D.) are shown in Table 1, below,
10 for those first-immune sera.

Table 1

HDL Levels In Immunized
Animals (mg/dl)

<u>Immunogen</u>	Avg. ³	<u>S.D.</u> ⁴	<u>P</u> ⁵
	<u>HDL</u>		
Control	23.89	3.92	—
rhCETP ¹	26.59	4.41	0.17
CETP-TH ²	26.14	6.93	0.38

15 ¹ rhCETP = Recombinant human CETP.

² CETP-TH = C-terminal 26 rabbit CETP amino acid residues conjugated to thyroglobulin.

20 ³ Avg. HDL = Average HDL concentration after immunization or mock immunization for the control.

⁴ S.D. = Standard deviation.

25 ⁵ P = p value from a Student's T test analysis.

As can be seen from those results, an increase in HDL particle concentration was found after administration of each of the CETP immunogens. There was a relatively large scatter in the data. Nevertheless, an approximately 10 percent increase in the HDL cholesterol level was observed with each CETP immunogen as compared with the control, with the recombinant human CETP immunogen providing its increase at a confidence level of greater than 80 percent (p=0.17) using a Student's T test to analyze the results.

Example 3: Construction Of *E. coli* Expression
Vectors Encoding HBcAg/CETP/HBcAg
Fusion Proteins

A. PCR amplification of HBcAg

Plasmid pFS14, a derivative of expression vector pKK223 (Pharmacia), encodes HBcAg (subtype ayw) [Schödel et al., *Infect. Immun.* 57:1347 (1989)]. PCR primer A, below, is designed to amplify the 5' end of HBcAg and place an NcoI (C'CATG,G) site in the correct reading frame at the natural ATG start codon. In each of the sequences shown hereinafter, only the coding strand is shown, and bases removed after cleavage by restriction enzymes are shown in lower case.

Primer A: 5' gatccCATGGACATCGACCCTTATAAAGAATTTGG 3'

SEQ ID NO:44

5 Primer Z: 5'gatcaAGCTTTTAACATTGAGATTCCCGAGAT
TGAGATCTTCTG 3' SEQ ID NO:45

15 The amplified DNA is then cleaved with NcoI and HindIII, and fractionated by size on an agarose gel. Full-length HBcAg DNA is purified from a gel slice using a QIAQUICK™ gel extraction kit (QIAGEN, Chatsworth, CA).

pProEx1, an *E. coli* expression vector (Life Technologies, Rockville, MD), is also cleaved with NcoI and HindIII and gel-purified. The amplified DNA and pProEx1 DNAs are ligated under standard conditions using T4 DNA ligase and transformed into chemically-competent *E. coli* DH10B cells (Life Technologies) using protocols supplied by the vendor to form plasmid ProEx1-AZ. The transformation mixture is spread on LB agar plates containing 100 μ g/ml ampicillin and incubated overnight at (about 18 hours) at 37°C. Colonies harboring ampicillin-resistant plasmids are purified by restreaking on fresh LB agar plates containing ampicillin, and minipreps of plasmid DNA are prepared

using WIZARD™ 373 DNA purification kits (Promega, Inc.,
Madison, WI). Plasmids containing the HBcAg fragment
inserted into the NcoI and HindIII sites of pProEx1 are
characterized by restriction mapping and sequence
5 analysis across the inserted region.

Plasmid pProEx1-AZ is then modified to insert
a polylinker between the nucleotides that encode amino
acid residues 70-75 of HBcAg.

Primer B is designed to insert an XhoI site
10 (C'TCGA) and an EcoRI site (G'AATT,C) site following
position 206 of SEQ ID NO:39. Primer Y is designed to
insert an EcoRI site (G'AATT,C) site followed by a SpeI
site (A'CTAG,T) before position 226 of SEQ ID NO:39.

15 Primer Y: 5' gatcgAATTCAGTTGGAAGATCCAGCGT
CTAGAGACCTAGTAG 3' SEQ ID NO:46

Primer B: 5' gatcgAATTCCTCGAGCTAGAGTCATTAGTT
20 CCCCCAGCA 3' SEQ ID NO:47

Plasmid pProEx1-AZ is then used as a template
with primers A and B to amplify a segment of DNA
(designated HBcAg-AB) encoding amino acid residues 1-69
of HBcAg to generate a fragment that contains an NcoI
25 site at its 5' end and an XhoI and a EcoRI site at its
3' end. The same plasmid is also used with primers Y
and Z to amplify a segment of DNA (designated HBcAg-YZ)
encoding amino acid residues 76-183 of HBcAg to generate
a fragment that contains EcoRI and SpeI sites at its 5'
30 end and a HindIII site at its 3' end.

The PCR product from the reaction designed to
produce plasmid HBcAg-AB is cleaved with NcoI and EcoRI
and purified after agarose gel electrophoresis. The PCR
product from a second reaction designed to produce
35 plasmid HBcAg-YZ is cleaved with EcoRI and HindIII and

purified after agarose gel electrophoresis. The two
gel-purified fragments are then ligated in a triple
ligation reaction to plasmid pProEx1 that had been
treated with NcoI and HindIII and purified after agarose
5 gel electrophoresis. The desired ligated plasmid,
pProEx1-AB-YZ, is obtained by screening
ampicillin-resistant colonies for plasmids that have the
predicted structure by restriction analysis, and is
confirmed by DNA sequencing across the whole HBcAg
10 region, particularly the A, BY, and Z junctions.

C. Cloning Of CETP Segment

Encoding SEQ ID NO:29

A stably transformed CHO cell line transfected
15 with human CETP cDNA [Wang et al., *J. Biol. Chem.*,
270:612-618 (1995); Wang et al., *J. Biol. Chem.*,
267:17487-17490 (1992)] provides CETP cDNA that is used
as a template to amplify a segment (nucleotides 1346 to
1431) of the CETP coding sequence (SEQ ID NO: 1) that
20 encodes the human peptide (SEQ ID NO:29;
ArgAspGlyPheLeuLeuLeuGlnMetAspPheGlyPheProGluHisLeu
LeuValAspPheLeuGlnSerLeuSer) that is bound by the
antibody TP2; T.L. Swenson et al., *J. Biol. Chem.*,
264:14318-14326 (1989).

25 Primer C, below, is designed to amplify a
region from just upstream from the natural XhoI site at
position 1346. Primer X, below, is designed to amplify
a region at the 3' end of the CETP gene, removing the
TAG codon and replacing it with an Eco47III site
30 (AGC|GCT) followed by an EcoRI site (G'AATT,C).

Primer C: 5' GATTATCACTCGAGATGGCTTCCTGCTGCTGCAG 3'

SEQ ID NO:48

Primer X: 5' gatcgAATTCAGCGCTCAAGCTCTGGAGG
AAATCCACCAG 3'

SEQ ID NO:49

5 The CETP cDNA is then used as a template with
primers C and X to amplify a segment of DNA (designated
pCETP-CX) encoding amino acid residues 461-476 of CETP,
that contains an XhoI site near its 5' end and an
Eco47III and EcoRI site at its 3' end. This segment,
CETP-CX, is then cleaved with XhoI and EcoR47III, and
10 gel-purified. Plasmid pProEx1-AB-YZ is digested with
SpeI and treated with T4 DNA polymerase to remove the
4-base 5' overhangs and generate blunt ends. [See, J.
Sambrook et al., Molecular Cloning, 2nd ed., Cold Spring
Harbor Press, Cold Spring Harbor N.Y., 8-23 (1989).]

15 The resulting plasmid is then treated with
XhoI, gel-purified, and ligated to the segment CETP-CX
that has an XhoI site at one end and a blunt end
resulting from cleavage with Eco47III at the other end.
The resulting plasmid, designated pProEx1-ABC-XYZ, is
20 characterized by restriction analysis and by sequencing
to confirm that it contains sequences encoding amino
acid residues 461-476 of CETP replacing sequences that
encoded amino acid residues 70-75 of HBcAg in the vector
pProEx1-AZ.

25

Example 4: Expression Of HBcAg/CETP/HbcAg
Fusion Proteins In *E. coli*

30 The pProEx1 vector is designed for the
expression of foreign proteins in *E. coli*. This vector
contains a gene conferring resistance to ampicillin and
a pBR322 origin of replication for propagation in *E.*
coli. It also has a multiple cloning site flanked by a
6 histidine sequence (6X His) and the recognition
sequence for rTEV protease. This site allows for the

removal of the 6X His tag from a fusion protein after purification. The vector also has a *Trc* promoter and *lacI^q* gene permitting inducible gene expression with isopropyl- β -D-thiogalactopyranoside (IPTG). A
5 procaryotic ribosomal binding site is located upstream from the start of translation of the 6X His tag. A unique *NcoI* site is located at the first codon of the 6X His tag. Plasmids pProEx1 and a control plasmid, pProEx1-CAT, are obtained from Life Technologies.

10 *E. coli* DH10B strains individually harboring pProEx1, pProEx1-CAT, pProEx1-AZ, or pProEx1-ABC-XYZ are cultured overnight (about 18 hours), and used as inocula for cultures that are induced with IPTG under conditions recommended by the vendor. Cultures harboring plasmid
15 pProEx1-AZ produce HBcAg and those harboring pProEx1-ABC-XYZ produce the desired HBcAg/CETP/HBcAg fusion protein as particles. These proteins lack the 6X His tag present in the original pProEx1 vector because the HBcAg sequences are inserted at the *NcoI* site at the
20 beginning of the 6X tag. Cultures harboring pProEx1-CAT produce a protein that migrates on SDS-PAGE gels as expected for a His-tagged CAT fusion protein.

25 Example 5: Expression Of HBcAg/CETP/HBcAg
Fusion Proteins In Baculovirus-
Infected Insect Cells

Baculovirus-infected insect cells have been shown to express a wide variety of recombinant proteins (V.A. Luckow, *Insect Cell Expression Technology*, pp.
30 183-218, in Protein Engineering: Principles and Practice, J.L. Cleland et al. eds., Wiley-Liss, Inc, 1996). Heterologous genes placed under the control of the polyhedrin promoter of the *Autographa californica* nuclear polyhedrosis virus (AcNPV) are often expressed

at high levels during the late stages of infection. In most cases, the recombinant proteins are appropriately processed and are functionally similar to their authentic counterparts.

5 Recombinant baculoviruses containing the chimeric HBcAg/CETP/HBcAg gene are constructed using the baculovirus shuttle vector system (Luckow et al., *J. Virol.*, 67:4566-4579, 1993) sold commercially as the Bac-To-Bac™ baculovirus expression system (Life
10 Technologies).

 Briefly, pProEx1-ABC-XYZ is digested with NcoI, treated with Klenow enzyme to fill in the ends, and digested with HindIII to release the entire fragment encoding the HBcAg/CETP/HBcAg fusion protein. This
15 fragment is inserted into a baculovirus donor plasmid, pFastBac1, that is digested with BamHI, treated with Klenow enzyme, and digested with HindIII. The resulting plasmid has the sequences encoding the hybrid
20 HBcAg/CETP/HBcAg gene inserted downstream from the polyhedrin promoter of AcNPV. The mini-Tn7 segment containing the polyhedrin/HBcAg/CETP/HBcAg expression cassette is then transposed to a baculovirus shuttle vector propagated in *E. coli* and colonies harboring
25 composite (recombinant) vectors are identified by their color and an altered drug resistance patterns. Miniprep DNAs are prepared and transfected into cultured *Spodoptera frugiperda* (fall armyworm) Sf9 cells.

Stocks of recombinant viruses are prepared and expression of the recombinant protein is monitored by standard protocols (O'Reilly et al., Baculovirus Expression Vectors: A Laboratory Manual, W.H. Freeman and Company, New York, 1992; King, L.A., and Possee, R.D. The Baculovirus Expression System: A Laboratory Guide, Chapman & Hall, London, 1992).

Example 6: Expression Of HBcAg/CETP/HBcAg
Fusion Proteins In Mammalian Cells

The HBcAg/CETP/HBcAg fusion protein is expressed in mammalian cell culture using the BHK/VP16 expression system (Hippenmeyer et al., *Bio/Technology*, 11:1037-1041, 1993). Briefly, the HbcAg/CETP/HbcAg-encoding sequence of the NcoI-HindIII fragment from plasmid pProEx1-ABC-XYZ of Example 3 is isolated by gel electrophoresis and purified as before. The fragment is treated with Klenow polymerase and all four nucleotide triphosphates to make the 5' overhanging ends blunt.

The mammalian expression vector pMON3327 contains the SV40 polyadenylation signal sequence in the BamHI site of plasmid pUC18, and is used as the basis for further plasmid construction. Ligation of the IE175 promoter of herpes simplex virus (HSV-1) upstream of the SV40 polyadenylation signal sequence in vector pMON3327 provides mammalian expression vector pMON3360B. The IE175 promoter is responsive to the HSV-1 VP-16 transactivator.

Expression vector pMON3360B is digested with BamHI and the 5' overhanging ends at the unique BamHI site are filled in using Klenow polymerase. The vector sequences and the HBcAg/CETP/HBcAg sequences are ligated overnight (about 18 hours) at 15°C using T4 DNA ligase. The ligation mixture is transfected into competent *E.*

coli and selected for ampicillin resistance. Plasmid DNA is isolated from the colonies and analyzed by restriction analysis for proper orientation of the HBcAg/CETP/HBcAg sequences in the pMON3360B vector. A plasmid with the correct orientation is designated pMON3360B-HBcAg-CETP. Plasmid pMON3360B-HBcAg-CETP is purified using Promega Maxiprep™ protocols from 400 ml cultures.

BHK/VP16 hamster kidney cells are plated at about 3×10^5 cells per 60 mm culture dish 24 hours before transfection in growth medium consisting of DMEM/5% fetal bovine sera (Life Technologies). Ten micrograms of plasmid pMON3360B-HBcAg-CETP and 1 μ g of plasmid pMON1118 are transfected into the cells using LipofectAmine™ (Life Technologies) as recommended by the manufacturer. Two days after transfection, the cells are treated with trypsin/EDTA (Life Technologies) and plated in ten 100 mm dishes in growth medium containing hygromycin (Sigma). In about two weeks, surviving colonies are isolated using filter paper and expanded and assayed for expression of the HBcAg/CETP/HBcAg fusion protein.

Example 7: Construction of DNA Vaccine Vectors
Capable of In Vivo Expression Of
HBcAg/CETP/HBcAg Fusion Proteins

A. Construction of pcDNAI/Amp-
HBcAg/CETP/HBcAg-1 and pcDNAI/Amp-
HBcAg/CETP/HBcAg-2

Vector pcDNAI/Amp (Invitrogen Corp., San Diego, CA) is a derivative of vector pcDNAI and its parent vector pCDM8. All three vectors have the following features: Enhancer-promoter sequences from the

immediate early gene of human cytomegalovirus (CMV) for high-level constitutive expression; SV40 poly(A) transcription termination and RNA processing signals to enhance mRNA stability, a versatile multiple cloning site to permit unidirectional or bidirectional cloning of inserts; and a ColE1 origin of replication for growth in *E. coli*. Vector pcDNAI/Amp also contains bacteriophage T7 and SP6 promoters flanking an expanded multiple cloning site, and an ampicillin resistance gene to facilitate growth and selection in most *E. coli* strains. These vectors can be used for high-level constitutive expression of recombinant proteins, including cytoplasmic proteins, transcription factors, viral proteins, cell surface receptors, and secreted proteins, in a variety of mammalian cells.

Vector pcDNAI/Amp can be linearized at any of a variety of positions in the multiple cloning site downstream from the pCMV promoter for insertion of heterologous genes. A blunt-ended DNA segment encoding HBcAg/CETP/HBcAg is prepared as described in Example 5 by digesting pProEx1-ABC-XYZ with NcoI and HindIII, and treating with Klenow enzyme in the presence of all four nucleotide triphosphates to make the 5' overhanging ends blunt. This fragment is purified from an agarose gel. Vector pcDNAI/Amp is digested with EcoRV to leave blunt ends, treated with shrimp alkaline phosphatase to remove 5' terminal phosphates, and ligated to the purified fragment encoding HBcAg/CETP/HBcAg. The resulting DNA is transformed into *E. coli* DH10B, and the resulting colonies are screened for plasmids containing the HBcAg/CETP/HBcAg inserted in the proper orientation downstream from the CMV promoter. The desired plasmid is designated pcDNAI/Amp-HBcAg/CETP/HBcAg-1.

A similar plasmid is also prepared by inserting the blunt-ended HBcAg/CETP/HBcAg into pcDNAI/Amp that is treated with BamHI, XbaI, and Klenow enzyme, to fill in the ends, to generate pcDNAI/Amp-
5 HBcAg/CETP/HBcAg-2. This plasmid differs from pcDNAI/Amp-HBcAg/CETP/HBcAg-1 by removal of the large central portion of the multiple cloning site flanked by the CMV promoter and the SV40 poly(A) regions.

10 B. Construction of pCMV-SPORT-HBcAg/CETP/HBcAg

Plasmid pCMV-SPORT- β -gal (Life Technologies, Rockville, MD) contains the *E. coli* β -galactosidase gene cloned as a NotI fragment into pCMV-SPORT1. These vectors contain a CMV promoter, an SV40 poly(A) site, an
15 ampicillin resistance gene, and an *E. coli* plasmid origin of replication. Plasmid pCMV-SPORT- β -gal is commonly used as a reporter vector to monitor transfection efficiency. The plasmid pCMV-SPORT-HBcAg/CETP/HBcAg is prepared using similar procedures
20 describe above in subsection A. Plasmid pCMV-SPORT- β -gal is linearized with NotI, treated with Klenow to fill in the ends, and with shrimp alkaline phosphatase to remove 5' terminal phosphates. The linearized, phosphatase-treated vector is then ligated to the blunt
25 end linear fragment from above encoding HBcAg/CETP/HBcAg. The ligation mixtures are transformed into *E. coli* DH10B, and the resulting plasmids screened for the proper orientation of the insert into the vector.

30 Plasmids pCMV-SPORT- β -gal and pGreen Lantern-1 (Life Technologies, Rockville, MD) both contain reporter genes under the control of the CMV promoter and are suitable as control vectors for use in mammalian transfection studies. Expression of β -galactosidase

activity is easily monitored by *in situ* staining of prokaryotic or eukaryotic cells with the chromogenic substrate X-gal. Plasmid pGreen Lantern-1 contains a mutated form of the gene for Green Fluorescent Protein (GFP) from *Aequorea victoria* jellyfish. GFP requires no substrates for visualization, and can be monitored in living as well as fixed cells, and in whole animals, by fluorescence microscopy using FITC filters.

10 Example 8: Preparation of purified DNA template
Briefly, cultures of *E. coli* harboring plasmids are grown up in TB or LB media. [See, J. Sambrook et al., Molecular Cloning, 2nd ed., Cold Spring Harbor Press, Cold Spring Harbor N.Y., 8-23 (1989).]
15 Plasmid DNA is released from pelleted cells by an alkaline lysis method using the reagents supplied in a commercial anion exchange purification kit (Qiagen GmbH, Hilden, Germany). The recovered lysate is purified on P-2500 columns (Qiagen), precipitated with isopropanol, solubilized in Tris-EDTA, reprecipitated in NaCl
20 ethanol, and finally resuspended in sterile endotoxin-free phosphate buffered saline (PBS; Sigma Chemical, St. Louis, MO). Plasmid DNA is stored at -20 degrees C. The bulk product is tested for the presence of sterility and the presence of contaminating DNA, RNA, protein, and
25 endotoxin according to established protocols [reviewed in M. Marquet et al., *BioPharm*, 10(5): 42-50 (1997)].

30 Example 9: In Vivo Expression of DNA Encoding HBcAg/CETP/HBcAg Fusion Proteins Injected Directly into the Muscles Of Mice

Endotoxin-free samples of plasmids pcDNAI/Amp-HBcAg/CETP/HBcAg-1 and -2, pCMV-SPORT-HBcAg/CETP/HBcAg,

pCMV-SPORT- β -gal, and pGreen Lantern-1 are prepared as described in the preceding examples. The quadriceps muscles of mice are injected with 100 μ g of one of the plasmids listed above, and the muscle tissue at the site of injection is assayed for activity of the β -gal or GFP reporter gene or expression of the HBcAg/CETP/HBcAg fusion protein after a suitable maintenance time.

A. Light microscopy

The quadriceps muscles of mice immunized by to the plasmid DNA injections, are removed in their entirety, cross-sectioned, and histochemically-stained with X-gal for β -gal activity [Wolff et al., *Science*, 247:1465-1468, 1990]. Only those tissues exposed to pCMV-SPORT- β -gal produce the insoluble precipitate that is the indigo blue product resulting from cleavage of X-gal by the expressed β -galactosidase protein. Similarly, samples expressing GFP when viewed by light microscopy using FITC filters, show a dramatic fluorescent signal with an excitation peak of 490 nm. [R. Heim, et al., *Nature*, 373: 663 (1995).] Samples expressing the fusion protein do not fluoresce under these conditions nor react with X-gal.

B. Immunofluorescence

Muscle samples expressing the HBcAg/CETP/HBcAg fusion protein are detected by immunofluorescence with primary antibodies directed against HBcAg or CETP, and any of a number of well-known secondary antibodies (e.g. FITC- or Rhodamine-conjugated secondary antibodies). Established immunofluorescence protocols are widely known in the art, and many of the reagents and methods are available from commercial sources.

C. Immunoblotting

Muscle extracts are prepared by mincing quadriceps tissue in a microcentrifuge tube containing lysis buffer (20 mM Tris, pH 7.4, 2 mM MgCl₂, and 0.1% Triton X-100) and grinding with a plastic pestle until homogenized. Protein samples are analyzed on SDS-PAGE gels and electrophoretically-transferred to Immobilon-P or nitrocellulose membranes, using standard protocols. [See, J. Sambrook et al., Molecular Cloning, 2nd ed., Cold Spring Harbor Press, Cold Spring Harbor N.Y., 8-23 (1989).]. Primary antibodies directed against β -galactosidase, GFP, HBcAg, and CETP obtained from commercial sources are used to detect the expression products resulting from injection of plasmid DNAs into the muscle tissue. Many secondary antibodies are available from a wide variety of commercial sources that are chemically conjugated to a reporter enzymes. Commercially available examples include alkaline phosphatase-conjugated anti-rabbit, anti-mouse, or anti-human IgG. Secondary antibodies chemically conjugated to horse radish peroxidase or to β -galactosidase are also widely available. Muscle extracts expressing GFP are detected with anti-GFP antibodies, those expressing β -gal detected with anti- β -gal antibodies, and those expressing the HBcAg/CETP/HBcAg fusion protein are detected with antibodies directed against HBcAg or CETP.

D. PCR analysis

Muscle extracts are also analyzed to detect presence of the injected plasmid DNAs by polymerase chain reaction techniques. Briefly, muscle tissues taken from the site of injection are homogenized in lysis buffer, template DNA is prepared, and used in reactions containing appropriate mixes of primers and a

thermostable polymerase. Primers lying within the
ampicillin resistance gene, common to all vectors,
amplify a DNA fragment in all tissues that take up and
maintain the injected plasmid DNA. Primer sets specific
5 for each gene, GFP, β -gal, HBcAg, and CETP, amplify
uniquely-sized fragments, if the plasmid insert is
intact within plasmids maintained within the cells. PCR
can be used to monitor the long term stability of the
plasmid within muscle tissues and aid in transcription
10 and expression studies, if expression levels monitored
by immunoblotting or microscopic techniques are low or
nonexistent. PCR can also be used to determine whether
the plasmids integrate into the host chromosome or are
lost by passive diffusion.

15

E. Antibody Production

Mice immunized as discussed above produce
antibodies that immunoreact with CETP in ELISA assays as
discussed before. Similarly immunized rabbits also
20 produce anti-CETP antibodies, which antibodies cause a
lessening in the transfer of cholesteryl esters from
HDL, and also an increase in the HDL particle
concentration in blood plasma.

25

Example 10: In Vivo Expression of DNA
Encoding Rabbit CETP Injected
Directly into the Muscles of Mice

30

A. PCR Amplification of Rabbit CETP

Rabbit CETP cDNA (SEQ ID NO: 27) [Nagashima et
al., J. Lipid Res., 29:1643-1649 (1988) or Kotake et
al., *Ibid*, 37:599-605 (1996)] is obtained as described.
PCR Primer N, below is designed to amplify the 5'end of
35 rabbit CETP and place an NotI (GC'GGCC,GC) and an NcoI

(C'CATG,G) site in the correct reading frame at the natural ATG start condon immediately preceding the GCC condon at position +1 in SEQ ID NO:27. In each of the sequences shown below, only the coding strand is shown, with the bases removed after cleavage by restriction enzymes not being shown, and synthetic sequences being shown in lower case.

Primer N:

10 5' ggccgccccatgGCCTGTCCCAAAGGCGCCTCCTACGAGGCT 3'

(SEQ ID NO:51)

15 Primer M, below, is designed to amplify the 3' end of the rabbit CETP and place a TAA stop condon, a HindIII (A'ACGT,T) and NotI site (GC'GGCC,GC) following the amino acid residue 497 (Ser) immediately preceding the TAG stop codon ending at position +1494 of SEQ ID NO:27

20

Primer M

5' ggccgcacgttttaCTAGCTCAGGCTCTGCAGGAAATCCACCAGCAGGTG

(SEQ ID NO:52)

25

30 A DNA fragment encoding the full-length rabbit CETP with modified 5' and 3' ends is amplified using the above cDNA as a template in the presence of primer M and primer N under the standard polymerase chain reaction conditions recommended by the manufacturer of the GeneAmp PCR reagent kit (Perkin Elmer Cetus, Norwalk, Conn.). The amplified DNA is then cleaved with NotI and fractionated by size on an agarose gel. Full-length rabbit CETP DNA is purified as described above.

35

B. Insertion of Rabbit CETP DNA into
pGEM -5Zf(+), pCR II, and pProEx1

5 The full-length rabbit CETP with NotI-compatible ends is inserted into a plasmid cloning vector, such as pGEMTM-5Zf(+) (Promega Corporation, Madison, WI), which has a unique NotI site in the polylinker region within the lacZalpha peptide region. Recombinants are selected by blue/white color screening
10 as recommended by the vendor. An alternate procedure is to clone the PCR product containing the full-length rabbit CETP directly into a TA cloning vector, such as pCRTM II (Invitrogen Corporation, San Diego, CA). DNA sequencing is carried out to confirm the identity of the
15 nucleotide sequence encoding the rabbit CETP. In either case, the full-length rabbit CETP DNA can be released from the vector as a fragment with NotI-compatible ends, or as an NcoI-HindIII fragment.

20 C. Expression of Rabbit CETP in *E. coli*,
Insect Cells, and Mammalian Cells

The rabbit CETP is inserted into pProEx1 as an NcoI-HindIII fragment as described above in Examples 3B
25 and 3C for the DNA encoding the HbcAg/CETP/HbcAg chimeric fusion protein. Expression of the rabbit CETP protein in *E. coli* is also carried out as described above in Example 4. The rabbit CETP protein is then purified from *E. coli*, to confirm the biological
30 activity of the isolated protein.

The DNA encoding rabbit CETP is purified and inserted into plasmid pFastBac1 as described in Example 5. Recombinant baculoviruses are generated and used to express rabbit CETP in infected insect cells.

The DNA encoding rabbit CETP is purified and inserted into pMON3327 as described in Example 6. BHK/VP16 cells are transfected with the resulting plasmid and surviving colonies are expanded and assayed for expression of the rabbit CETP.

D. Insertion of the Rabbit CETP gene
into DNA Vaccine Vectors

The rabbit CETP gene (rCETP) is inserted as a blunt-ended fragment into vector pcDNAI/Amp as described in Example 7A. The rabbit CETP gene is also inserted into pCMV-SPORT as an NotI fragment as described in Example 7B. Both plasmids direct expression of rabbit CETP in mammalian cells under the control of the CMV promoter.

Mammalian cells are transfected with pcDNAI/Amp-rCETP or pCMV-SPORT-rCETP. Expression of rabbit CETP is assayed by immunofluorescence techniques as described in Example 9B and by immunoblotting techniques, as described in Example 9C.

Once rabbit CETP is detectable and expression levels are suitably high in transfected mammalian cell lines to provide assurance of expression, injection of pcDNAI/Amp-rCETP or pCMV-SPORT-rCETP into muscle tissues of animals containing CETP in their blood, such as Guinea pigs for example, is carried out and monitored as described in Examples 9A-E.

The foregoing description and the examples are intended as illustrative and are not to be taken as limiting. Still other variations within the spirit and scope of this invention are possible and will readily present themselves to those skilled in the art.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Needleman, Philip
Glenn, Kevin
- (ii) TITLE OF INVENTION: An Immunological Process and Constructs
for Increasing the HDL Cholesterol Concentration by DNA
Vaccination
- (iii) NUMBER OF SEQUENCES: 52
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Welsh & Katz, Ltd.
 - (B) STREET: 120 South Riverside Plaza, 22nd Floor
 - (C) CITY: Chicago
 - (D) STATE: IL
 - (E) COUNTRY: USA
 - (F) ZIP: 60606
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Gamson Ph.D., Edward P.
 - (B) REGISTRATION NUMBER: 29,381
 - (C) REFERENCE/DOCKET NUMBER: MON-103.0 6221/69666
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (312)655-1500
 - (B) TELEFAX: (312)655-1501

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1431 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (viii) POSITION IN GENOME:
 - (C) UNITS: bp
- (x) PUBLICATION INFORMATION:
 - (A) AUTHORS: Drayna, Dennis
Jarnagin, Alisha Stephens
McLean, John
Henzel, William
Kohr, William
Fielding, Christopher
Lawn, Richard

(B) TITLE: Cloning and sequencing of human cholesteryl
ester transfer protein cDNA
(C) JOURNAL: Nature
(D) VOLUME: 327
(F) PAGES: 632-634
(G) DATE: June 18-1987

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TGCTCCAAAG GCACCTCGCA CGAGGCAGGC ATCGTGTGCC GCATCACCAA GCCTGCCCTC	60
CTGGTGTGTA ACCACGAGAC TGCCAAGGTC ATCCAGACCG CCTTCCAGCG AGCCAGCTAC	120
CCAGATATCA CGGGCGAGAA GGCCATGATG CTCCTTGGCC AAGTCAAGTA TGGGTTGCAC	180
AACATCCAGA TCAGCCACTT GTCCATCGCC AGCAGCCAGG TGGAGCTGGT GGAAGCCAAG	240
TCCATTGATG TCTCCATTCA GAACGTGTCT GTGGTCTTCA AGGGGACCCT GAAGTATGGC	300
TACACCACTG CCTGGTGGCT GGGTATTGAT CAGTCCATTG ACTTCGAGAT CGACTCTGCC	360
ATTGACCTCC AGATCAACAC ACAGCTGACC TGTGACTCTG GTAGAGTGCG GACCGATGCC	420
CCTGACTGCT ACCTGTCTTT CCATAAGCTG CTCCTGCATC TCCAAGGGGA GCGAGAGCCT	480
GGGTGGATCA AGCAGCTGTT CACAAATTTC ATCTCCTTCA CCCTGAAGCT GGTCTGAAG	540
GGACAGATCT GCAAAGAGAT CAACGTCATC TCTAACATCA TGGCCGATTT TGTCCAGACA	600
AGGGGTGCCA GCATCCTTTC AGATGGAGAC ATTGGGGTGG ACATTTCCTT GACAGGTGAT	660
CCCCGTCATCA CAGCCTCCTA CCTGGAGTCC CATCACAAGG GTCATTTTCT CTACAAGAAT	720
GTCTCAGAGG ACCTCCCCCT CCCCACCTTC TCGCCACAC TGCTGGGGGA CTCCCGCATG	780
CTGTACTTCT GGTCTCTGTA GCGAGTCTTC CACTCGCTGG CCAAGGTAGC TTTCCAGGAT	840
GGCCGCCTCA TGCTCAGCCT GATGGGAGAC GAGTTCAAGG CAGTGCTGGA GACCTGGGGC	900
TTCAACACCA ACCAGGAAAT CTTCCAAGAG GTTGTGCGCG GCTTCCCCAG CCAGGCCCAA	960
GTCACCGTCC ACTGCCTCAA GATGCCCAAG ATCTCCTGCC AAAACAAGGG AGTCGTGGTC	1020
AATTCTTCAG TGATGGTGAA ATTCTCTTT CCACGCCAG ACCAGCAACA TTCTGTAGCT	1080
TACACATTTG AAGAGGATAT CGTGACTACC GTCCAGGCCT CCTATTCTAA GAAAAAGCTC	1140
TTCTTAAGCC TCTTGGATTT CCAGATTACA CCAAAGACTG TTTCCAACCT GACTGAGAGC	1200
AGCTCCGAGT CCATCCAGAG CTTCTGTCAG TCAATGATCA CCGCTGTGGG CATCCCTGAG	1260
GTCATGTCTC GGCTCGAGGT AGTGTTTACA GCCCTCATGA ACAGCAAAGG CGTGAGCCTC	1320
TTGACATCA TCAACCCTGA GATTATCACT CGAGATGGCT TCCTGCTGCT GCAGATGGAC	1380
TTTGGCTTCC CTGAGCACCT GCTGGTGGAT TTCCTCCAGA GCTTGAGCTA G	1431

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Glu Ile Phe Gln Glu Leu Ser Arg Gly Leu Pro Thr Gly Gln Ala Gln
1 5 10 15
Val Ala Val His
20

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Val Ala Val Thr Phe Arg Phe Pro Arg Pro Asp Gly Arg Glu Ala Val
1 5 10 15
Ala Tyr Arg Phe
20

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Leu Leu Leu Gln Met Asp Phe Gly Phe Pro Lys His Leu Leu Val Asp
1 5 10 15
Phe Leu Gln Ser Leu Ser
20

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Thr Thr Val Gln Ala Ser Tyr Ser Gln Lys Lys Leu Phe Leu His Leu
1 5 10 15
Leu Asp Phe Gln
20

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Leu Leu Leu His Leu Gln Gly Glu Arg Glu Pro Gly Trp Leu Lys Gln
1 5 10 15
Leu Phe Thr Asn
 20

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Asp Val Ser Gly Glu Arg Ala Val Met Leu Leu Gly Arg Val Lys Tyr
1 5 10 15
Gly Leu His Asn
 20

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Gln Glu Ile Phe Gln Glu Val Val Gly Gly Phe Pro Ser Gln Ala Gln
1 5 10 15
Val Thr Val His
 20

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Val Met Val Lys Phe Leu Phe Pro Arg Pro Asp Gln Gln His Ser Val
1 5 10 15
Ala Tyr Thr Phe
20

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Leu Leu Leu Gln Met Asp Phe Gly Phe Pro Glu His Leu Leu Val Asp
1 5 10 15
Phe Leu Gln Ser Leu Ser
20

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Thr Thr Val Gln Ala Ser Tyr Ser Lys Lys Lys Leu Phe Leu Ser Leu
1 5 10 15
Leu Asp Phe Gln
20

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Leu Leu Leu His Leu Gln Gly Glu Arg Glu Pro Gly Trp Ile Lys Gln
 1 5 10 15
 Leu Phe Thr Asn
 20

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Asp Ile Thr Gly Glu Lys Ala Met Met Leu Leu Gly Gln Val Lys Tyr
 1 5 10 15
 Gly Leu His Asn
 20

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 63 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
 (C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

CAGGAAATCT TCCAGGAGCT TTCCAGAGGC CTTCCACCG GCCAGGCCCA GGTAGCCGTC 60
 CAC 63

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 60 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
 (C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GTCGCCGTGA CGTTCGCTT CCCCCGCCCA GATGGCCGAG AAGCTGTGGC CTACAGGTTT 60

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 66 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CTGCTGCTGC AGATGGACTT CGGTTTTCCC AAGCACCTGC TGGTGGATTT CCTGCAGAGC 60
CTGAGC 66

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ACCACCGTCC AGGCCTCCTA CTCCCAGAAA AAGCTCTTCC TACACCTCTT GGATTTCAG 60

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTGCTCCTGC ACCTCCAGGG GGAGCGCGAG CCGGGGTGGC TCAAGCAGCT CTTACAAAC 60

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GACGTCAGCG GCGAGAGGGC CGTGATGCTC CTCGGCCGGG TCAAGTACGG GCTGCACAAC 60

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 63 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CAGGAAATCT TCCAAGAGGT TGTCGGCGGC TTCCCCAGCC AGGCCCAAGT CACCGTCCAC 60
TGC 63

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

GTGATGGTGA AATTCCTCTT TCCACGCCCA GACCAGCAAC ATTCTGTAGC TTACACATTT 60

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 66 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (viii) POSITION IN GENOME:
 - (C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

CTGCTGCTGC AGATGGACTT TGGCTTCCCT GAGCACCTGC TGGTGGATTT CCTCCAGAGC 60
TTGAGC 66

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (viii) POSITION IN GENOME:
 - (C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

ACTACCGTCC AGGCCTCCTA TTCTAAGAAA AAGCTCTTCT TAAGCCTCTT GGATTTCCAG 60

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (viii) POSITION IN GENOME:
 - (C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CTGCTCCTGC ATCTCCAAGG GGAGCGAGAG CCTGGGTGGA TCAAGCAGCT GTTCACAAAT 60

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
(C) UNITS: bp

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GATATCACGG GCGAGAAGGC CATGATGCTC CTTGGCCAAG TCAAGTATGG GTTGACACAAC 60

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 497 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(x) PUBLICATION INFORMATION:

- (A) AUTHORS: Nagashima, M.
McLean, J. W.
Lawn, R. M.
- (B) TITLE: Cloning and mRNA tissue distribution of
rabbit cholesteryl ester transfer protein
- (C) JOURNAL: J. Lipid Res.
- (D) VOLUME: 29
- (F) PAGES: 1643-1649
- (G) DATE: 1988

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Ala	Cys	Pro	Lys	Gly	Ala	Ser	Tyr	Glu	Ala	Gly	Ile	Val	Cys	Arg	Ile
1				5				10					15		
Thr	Lys	Pro	Ala	Leu	Leu	Val	Leu	Asn	Gln	Glu	Thr	Ala	Lys	Val	Val
			20					25					30		
Gln	Thr	Ala	Phe	Gln	Arg	Ala	Gly	Tyr	Pro	Asp	Val	Ser	Gly	Glu	Arg
			35				40					45			
Ala	Val	Met	Leu	Leu	Gly	Arg	Val	Lys	Tyr	Gly	Leu	His	Asn	Leu	Gln
			50			55					60				
Ile	Ser	His	Leu	Ser	Ile	Ala	Ser	Ser	Gln	Val	Glu	Leu	Val	Asp	Ala
65					70					75				80	
Lys	Thr	Ile	Asp	Val	Ala	Ile	Gln	Asn	Val	Ser	Val	Val	Phe	Lys	Gly
				85					90					95	
Thr	Leu	Asn	Tyr	Ser	Tyr	Thr	Ser	Ala	Trp	Gly	Leu	Gly	Ile	Asn	Gln
			100					105					110		
Ser	Val	Asp	Phe	Glu	Ile	Asp	Ser	Ala	Ile	Asp	Leu	Gln	Ile	Asn	Thr
			115				120					125			

[illegible]

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1494 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(viii) POSITION IN GENOME:
 (C) UNITS: bp

- (x) PUBLICATION INFORMATION:
 (A) AUTHORS: Nagashima, Mariko
 McLean, John W.
 Lawn, Richard M.
 (B) TITLE: Cloning and mRNA tissue distribution of
 rabbit cholesteryl ester transfer protein
 (C) JOURNAL: J. Lipid Res.
 (D) VOLUME: 29
 (F) PAGES: 1643-1649
 (G) DATE: 1988

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

GCCTGTCCCA AAGGCGCCTC CTACGAGGCT GGCATCGTGT GTCGCATCAC CAAGCCCCGCC	60
CTCTTGGTGT TGAACCAAGA GACGGCCAAG GTGGTCCAGA CGGCCTTCCA GCGCGCCGGC	120
TATCCGGACG TCAGCGGCGA GAGGGCCGTG ATGCTCCTCG GCCGGGTCAA GTACGGGCTG	180
CACAACCTCC AGATCAGCCA CCTGTCCATC GCCAGCAGCC AGGTGGAGCT GGTGGACGCC	240
AAGACCATCG ACGTCGCCAT CCAGAACGTG TCCGTGGTCT TCAAGGGGAC CCTGAACCTAC	300
AGCTACACGA GTGCCTGGGG GTTGGGCATC AATCAGTCTG TCGACTTCGA GATCGACTCT	360
GCCATTGACC TCCAGATCAA CACAGAGCTG ACCTGCGACG CTGGCAGTGT GCGCACCAAT	420
GCCCCGACT GCTACCTGGC TTTCCATAAA CTGCTCCTGC ACCTCCAGGG GGAGCGCGAG	480
CCGGGGTGGC TCAAGCAGCT CTTCAAAAC TTCATCTCCT TCACCCTGAA GCTGATTCTG	540
AAGCGACAGG TCTGCAATGA GATCAACACC ATCTCCAACA TCATGGCTGA CTTTGTCCAG	600
ACGAGGGCCG CCAGCATCCT CTCAGATGGA GACATCGGGG TGGACATTTC CGTGACGGGG	660
GCCCCGTGCA TCACAGCCAC CTACCTGGAG TCCCATCACA AGGGTCACTT CACGCACAAG	720
AACGTCTCCG AGGCCTTCCC CCTCCGCGCC TTCCCGCCCG GTCTTCTGGG GGACTCCCGC	780
ATGCTCTACT TCTGGTTCTC CGATCAAGTG CTCAACTCCC TGGCCAGGGC CGCCTTCCAG	840
GAGGGCCGTC TCGTGCTCAG CCTGACAGGG GATGAGTTCA AGAAAGTGCT GGAGACCCAG	900
GGTTTCGACA CCAACCAGGA AATCTTCCAG GAGCTTTCCA GAGGCCTTCC CACCGGCCAG	960
GCCCAGGTAG CCGTCCACTG CCTTAAGGTG CCCAAGATCT CCTGCCAGAA CCGGGGTGTC	1020
GTGGTGTCTT CTTCCGTCGC CGTGACGTTT CGCTTCCCCC GCCCAGATGG CCGAGAAGCT	1080
GTGGCCTACA GGTTCGAGGA GGATATCATC ACCACCGTCC AGGCCTCCTA CTCCAGAAA	1140
AAGCTCTTCC TACACCTCTT GGATTTCAG TCGTGCCGG CCAGCGGAAG GGCAGGCAGC	1200

TCAGCAAATC TCTCCGTGGC CCTCAGGACT GAGGCTAAGG CTGTTTCCAA CCTGACTGAG 1260
 AGCCGCTCCG AGTCCCTGCA GAGCTCTCTC CGCTCCCTGA TCGCCACGGT GGGCATCCCG 1320
 GAGGTTCATGT CTCGGCTCGA GGTGGCGTTC ACAGCCCTCA TGAACAGCAA AGGCCTGGAC 1380
 CTCTTCGAAA TCATCAACCC CGAGATTATC ACTCTCGATG GCTGCCTGCT GCTGCAGATG 1440
 GACTTCGGTT TTCCCAAGCA CCTGCTGGTG GATTTCTGCG AGAGCCTGAG CTAG 1494

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 476 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(x) PUBLICATION INFORMATION:

- (A) AUTHORS: Drayna, Dennis
 Jarnagin, Alisha Stephens
 McLean, John
 Henzel, William
 Kohr, William
 Fielding, Christopher
 Lawn, Richard
- (B) TITLE: Cloning and sequencing of human cholesteryl
 ester transfer protein cDNA
- (C) JOURNAL: Nature
- (D) VOLUME: 327
- (F) PAGES: 632-634
- (G) DATE: June 18-1987

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Cys	Ser	Lys	Gly	Thr	Ser	His	Glu	Ala	Gly	Ile	Val	Cys	Arg	Ile	Thr
1				5					10					15	
Lys	Pro	Ala	Leu	Leu	Val	Leu	Asn	His	Glu	Thr	Ala	Lys	Val	Ile	Gln
			20					25					30		
Thr	Ala	Phe	Gln	Arg	Ala	Ser	Tyr	Pro	Asp	Ile	Thr	Gly	Glu	Lys	Ala
		35					40					45			
Met	Met	Leu	Leu	Gly	Gln	Val	Lys	Tyr	Gly	Leu	His	Asn	Ile	Gln	Ile
	50					55					60				
Ser	His	Leu	Ser	Ile	Ala	Ser	Ser	Gln	Val	Glu	Leu	Val	Glu	Ala	Lys
65					70					75					80
Ser	Ile	Asp	Val	Ser	Ile	Gln	Asn	Val	Ser	Val	Val	Phe	Lys	Gly	Thr
			85						90					95	
Leu	Lys	Tyr	Gly	Tyr	Thr	Thr	Ala	Trp	Trp	Leu	Gly	Ile	Asp	Gln	Ser
			100					105					110		
Ile	Asp	Phe	Glu	Ile	Asp	Ser	Ala	Ile	Asp	Leu	Gln	Ile	Asn	Thr	Gln
		115					120					125			
Leu	Thr	Cys	Asp	Ser	Gly	Arg	Val	Arg	Thr	Asp	Ala	Pro	Asp	Cys	Tyr
		130				135					140				
Leu	Ser	Phe	His	Lys	Leu	Leu	Leu	His	Leu	Gln	Gly	Glu	Arg	Glu	Pro
145					150					155					160

Gly	Trp	Ile	Lys	Gln	Leu	Phe	Thr	Asn	Phe	Ile	Ser	Phe	Thr	Leu	Lys	165	170	175
Leu	Val	Leu	Lys	Gly	Gln	Ile	Cys	Lys	Glu	Ile	Asn	Val	Ile	Ser	Asn	180	185	190
Ile	Met	Ala	Asp	Phe	Val	Gln	Thr	Arg	Ala	Ala	Ser	Ile	Leu	Ser	Asp	195	200	205
Gly	Asp	Ile	Gly	Val	Asp	Ile	Ser	Leu	Thr	Gly	Asp	Pro	Val	Ile	Thr	210	215	220
Ala	Ser	Tyr	Leu	Glu	Ser	His	His	Lys	Gly	His	Phe	Ile	Tyr	Lys	Asn	225	230	235
Val	Ser	Glu	Asp	Leu	Pro	Leu	Pro	Thr	Phe	Ser	Pro	Thr	Leu	Leu	Gly	245	250	255
Asp	Ser	Arg	Met	Leu	Tyr	Phe	Trp	Phe	Ser	Glu	Arg	Val	Phe	His	Ser	260	265	270
Leu	Ala	Lys	Val	Ala	Phe	Gln	Asp	Gly	Arg	Leu	Met	Leu	Ser	Leu	Met	275	280	285
Gly	Asp	Glu	Phe	Lys	Ala	Val	Leu	Glu	Thr	Trp	Gly	Phe	Asn	Thr	Asn	290	295	300
Gln	Glu	Ile	Phe	Gln	Glu	Val	Val	Gly	Gly	Phe	Pro	Ser	Gln	Ala	Gln	305	310	315
Val	Thr	Val	His	Cys	Leu	Lys	Met	Pro	Lys	Ile	Ser	Cys	Gln	Asn	Lys	325	330	335
Gly	Val	Val	Val	Asn	Ser	Ser	Val	Met	Val	Lys	Phe	Leu	Phe	Pro	Arg	340	345	350
Pro	Asp	Gln	Gln	His	Ser	Val	Ala	Tyr	Tyr	Phe	Glu	Glu	Asp	Ile	Val	355	360	365
Thr	Thr	Val	Gln	Ala	Ser	Tyr	Ser	Lys	Lys	Lys	Leu	Phe	Leu	Ser	Leu	370	375	380
Leu	Asp	Phe	Gln	Ile	Thr	Pro	Lys	Thr	Val	Ser	Asn	Leu	Thr	Glu	Ser	385	390	395
Ser	Ser	Glu	Ser	Ile	Gln	Ser	Phe	Leu	Gln	Ser	Met	Ile	Thr	Ala	Val	405	410	415
Gly	Ile	Pro	Glu	Val	Met	Ser	Arg	Leu	Glu	Val	Val	Phe	Thr	Ala	Leu	420	425	430
Met	Asn	Ser	Lys	Gly	Val	Ser	Leu	Phe	Asp	Ile	Ile	Asn	Pro	Glu	Ile	435	440	445
Ile	Thr	Arg	Asp	Gly	Phe	Leu	Leu	Leu	Gln	Met	Asp	Phe	Gly	Phe	Pro	450	455	460
Glu	His	Leu	Leu	Val	Asp	Phe	Leu	Gln	Ser	Leu	Ser					465	470	475

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide

(x) PUBLICATION INFORMATION:
 (A) AUTHORS: Swenson, T. L.
 et al.,
 (C) JOURNAL: J. Biol. Chem.
 (D) VOLUME: 264
 (F) PAGES: 14318-14326
 (G) DATE: 1989

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Arg Asp Gly Phe Leu Leu Leu Gln Met Asp Phe Gly Phe Pro Glu His
 1 5 10 15
 Leu Leu Val Asp Phe Leu Gln Ser Leu Ser
 20 25

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 493 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(x) PUBLICATION INFORMATION:
 (A) AUTHORS: Pape, Michael E.
 Rehberg, Edward F.
 Marotti, Keith R.
 Melchior, George W.
 (B) TITLE: Molecular Cloning, Sequence, and Expression
 of Cynomolgus Monkey Cholesteryl Ester Transfer
 Protein
 (C) JOURNAL: Arteriosclerosis and Thrombosis
 (D) VOLUME: 11
 (E) ISSUE: 6
 (F) PAGES: 1759-1771
 (G) DATE: Nov/Dec-1991

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Met Leu Ala Ala Thr Val Leu Thr Leu Ala Leu Leu Gly Asn Val His
 1 5 10 15
 Ala Cys Ser Lys Gly Thr Ser His Lys Ala Gly Ile Val Cys Arg Ile
 20 25 30
 Thr Lys Pro Ala Leu Leu Val Leu Asn Gln Glu Thr Ala Lys Val Ile
 35 40 45
 Gln Ser Ala Phe Gln Arg Ala Asn Tyr Pro Asn Ile Thr Gly Glu Lys
 50 55 60
 Ala Met Met Leu Leu Gly Gln Val Lys Tyr Gly Leu His Asn Ile Gln
 65 70 75 80
 Ile Ser His Leu Ser Ile Ala Ser Ser Arg Val Glu Leu Val Glu Ala
 85 90 95
 Lys Ser Ile Asp Val Ser Ile Gln Asn Val Ser Val Val Phe Lys Gly
 100 105 110
 Thr Leu Lys Tyr Gly Tyr Thr Thr Ala Trp Gly Leu Gly Ile Asp Gln
 115 120 125

Ser	Val	Asp	Phe	Glu	Ile	Asp	Ser	Ala	Ile	Asp	Leu	Gln	Ile	Asn	Thr
130						135					140				
Gln	Leu	Thr	Cys	Asp	Ser	Gly	Arg	Val	Arg	Thr	Asp	Ala	Pro	Asp	Cys
145					150					155					160
Tyr	Leu	Ser	Phe	His	Lys	Leu	Leu	Leu	His	Leu	Gln	Gly	Glu	Arg	Glu
				165					170					175	
Pro	Gly	Trp	Ile	Lys	Gln	Leu	Phe	Thr	Asn	Phe	Ile	Ser	Phe	Thr	Leu
			180					185					190		
Lys	Leu	Val	Leu	Lys	Gly	Gln	Ile	Cys	Lys	Glu	Ile	Asn	Ile	Ile	Ser
		195					200					205			
Asn	Ile	Met	Ala	Asp	Phe	Val	Gln	Thr	Arg	Ala	Ala	Ser	Ile	Leu	Ser
	210					215					220				
Asp	Gly	Asp	Ile	Gly	Val	Asp	Ile	Ser	Leu	Thr	Gly	Asp	Pro	Ile	Ile
225					230						235				240
Thr	Ala	Ser	Tyr	Leu	Glu	Ser	His	His	Lys	Gly	Tyr	Phe	Ile	Tyr	Lys
				245					250					255	
Asn	Val	Ser	Glu	Asp	Leu	Pro	Leu	Pro	Thr	Phe	Ser	Pro	Ala	Leu	Leu
			260					265					270		
Gly	Asp	Ser	Arg	Met	Leu	Tyr	Phe	Trp	Phe	Ser	Glu	Gln	Val	Phe	His
		275					280					285			
Ser	Leu	Ala	Lys	Val	Ala	Phe	Gln	Asp	Gly	Arg	Leu	Thr	Leu	Ser	Leu
	290					295					300				
Met	Gly	Asp	Glu	Phe	Lys	Ala	Val	Leu	Glu	Thr	Trp	Gly	Phe	Asn	Thr
305					310					315					320
Asn	Gln	Glu	Ile	Phe	Gln	Glu	Val	Val	Gly	Gly	Phe	Pro	Ser	Gln	Ala
				325					330					335	
Gln	Val	Thr	Val	His	Cys	Leu	Lys	Met	Pro	Arg	Ile	Ser	Cys	Gln	Asn
			340					345					350		
Lys	Gly	Val	Val	Val	Asn	Ser	Ser	Val	Met	Val	Lys	Phe	Leu	Phe	Pro
		355					360					365			
Arg	Pro	Asp	Gln	Gln	His	Ser	Val	Ala	Tyr	Thr	Phe	Glu	Glu	Asp	Ile
	370					375					380				
Met	Thr	Thr	Val	Gln	Ala	Ser	Tyr	Ser	Lys	Lys	Lys	Leu	Phe	Leu	Ser
385					390					395					400
Leu	Leu	Asp	Phe	Gln	Ile	Thr	Pro	Lys	Thr	Val	Ser	Asn	Leu	Thr	Glu
				405					410					415	
Ser	Ser	Ser	Glu	Ser	Val	Gln	Ser	Phe	Leu	Gln	Ser	Met	Ile	Thr	Thr
			420					425					430		
Val	Gly	Ile	Pro	Glu	Val	Met	Ser	Arg	Leu	Glu	Ala	Val	Phe	Thr	Ala
		435					440					445			
Leu	Met	Asn	Ser	Lys	Gly	Leu	Ser	Leu	Phe	Asp	Ile	Ile	Asn	Pro	Glu
	450					455					460				
Ile	Ile	Thr	Arg	Asp	Gly	Phe	Leu	Leu	Leu	Gln	Met	Asp	Phe	Gly	Phe
465					470					475					480
Pro	Glu	His	Leu	Leu	Val	Asp	Phe	Leu	Gln	Ser	Leu	Ser			
				485					490						

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1508 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) PUBLICATION INFORMATION:

- (A) AUTHORS: Pape, Michael E.
Rehberg, Edward F.
Marotti, Keith R.
Melchior, George W.
- (B) TITLE: Molecular Cloning, Sequence, and Expression
of Cynomolgus Monkey Cholesteryl Ester Transfer
Protein
- (C) JOURNAL: Arteriosclerosis and Thrombosis
- (D) VOLUME: 11
- (E) ISSUE: 6
- (F) PAGES: 1759-1771
- (G) DATE: Nov/Dec-1991

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

ATGCTGGCTG CCACCGTCCT GACCCTGGCC CTGCTGGGCA ATGTCCACGC CTGCTCCAAA	60
GGTACCTCAC ACAAGGCAGG CATTGTGTGC CGCATCACCA AGCCTGCCCT CCTGGTGTTC	120
AACCAACAGA CTGCCAAGGT GATCCAGTCT GCCTTCCAGC GAGCCAACTA CCCAAATATC	180
ACAGGCGAGA AGGCCATGAT GCTCCTTGGC CAAGTCAAGT ATGGGTTGCA CAACATCCAA	240
ATCAGCCACT TGTCCATCGC CAGCAGCCGG GTGGAGCTGG TGGAAGCCAA GTCCATTGAT	300
GTCTCCATTC AGAACGTGTC TGTGGTCTTC AAGGGGACCC TGAAGTATGG CTACACCACT	360
GCCTGGGGGC TGGGCATTGA TCAGTCCGTT GACTTCGAGA TCGACTCTGC CATTGACCTC	420
CAGATCAACA CACAACGAC CTGTGACTCT GGTAGAGTGA GGACTGATGC CCCTGACTGC	480
TACCTGTCTT TCCATAAGCT GCTCCTGCAT CTCCAAGGGG AGCGAGAGCC CGGGTGGATC	540
AAGCAGCTGT TCACAACTT CATCTCCTTC ACCCTGAAGC TGGTCCTGAA GGGACAGATC	600
TGCAAAGAGA TCAACATCAT CTCCAACATC ATGGCCGATT TTGTCCAGAC AAGGGCTGCC	660
AGTATCCTTT CAGATGGAGA CATCGGGGTG GACATTTCCT TGACAGGTGA TCCCATCATT	720
ACAGCCTCCT ACCTGGAGTC CCATCACAAG GGTATTATTC TCTATAAGAA TGTCTCGGAG	780
GACCTCCCAC TCCCCACCTT CTCGCCCCGA CTGCTGGGGG ACTCCCGCAT GCTGTACTTC	840
TGGTTCTCCG AGCAAGTCTT CCACTCCCTG GCCAAGGTAG CTTTCCAAGA TGCCCGCCTC	900
ACGCTCAGCC TGATGGGAGA CGAGTTCAAG GCAGTGCTGG AGACCTGGGG CTTCAACACC	960
AACCAAGAAA TCTTCCAGGA GGTGTGCGGC GGCTTCCCCA GCCAGGCCCA AGTCACCGTC	1020
CACTGCCTCA AGATGCCCAG GATCTCCTGC CAAAACAAGG GAGTCGTGGT CAATTCTTCG	1080
GTGATGGTGA AATTCTCTT TCCACGCCCA GACCAGCAAC ACTCTGTAGC TTACACATTT	1140
GAAGAGGATA TCATGACCAC CGTCCAGGCC TCCTATTCTA AGAAAAAGCT CTTCTTAAGC	1200
CTCTTGATT TCCAGATTAC ACCAAAGACT GTTTCCTCACT TGACTGAGAG CAGCTCCGAG	1260

TCCGTCCAGA GCTTCCTGCA GTCAATGATC ACCACTGTGG GCATCCCTGA GGTTCATGTCT 1320
 CGGCTTGAGG CAGTGTTTAC AGCCCTCATG AACAGCAAAG GCCTGAGCCT CTTGACATC 1380
 ATCAATCCTG AGATTATCAC TCGAGATGGC TTCCTGCTGC TGCAGATGGA CTTTGGCTTC 1440
 CCTGAGCACC TGCTGGTGGG TTTCTCCAG AGCTTGAGCT AGAAGTCTCC AAGGACGTCA 1500
 GGATGGGG 1508

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Gln Glu Ile Phe Gln Glu Val Val Gly Gly Phe Pro Ser Gln Ala Gln
 1 5 10 15
 Val Thr Val His
 20

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Val Met Val Lys Phe Leu Phe Pro Arg Pro Asp Gln Gln His Ser Val
 1 5 10 15
 Ala Tyr Thr Phe
 20

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Leu Leu Leu Gln Met Asp Phe Gly Phe Pro Glu His Leu Leu Val Asp
 1 5 10 15

Phe Leu Gln Ser Leu Ser
20

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Thr Thr Val Gln Ala Ser Tyr Ser Lys Lys Lys Leu Phe Leu Ser Leu
1 5 10 15
Leu Asp Phe Gln
20

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Leu Leu Leu His Leu Gln Gly Glu Arg Glu Pro Gly Trp Ile Lys Gln
1 5 10 15
Leu Phe Thr Asn
20

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

Asn Ile Thr Gly Glu Lys Ala Met Met Leu Leu Gly Gln Val Lys Tyr
1 5 10 15
Gly Leu His Asn
20

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 183 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

```

Met Asp Ile Asp Pro Tyr Lys Glu Phe Gly Ala Thr Val Glu Leu Leu
 1      5      10      15
Ser Phe Leu Pro Ser Asp Phe Phe Pro Ser Val Arg Asp Leu Leu Asp
      20      25      30
Thr Ala Ser Ala Leu Tyr Arg Glu Ala Leu Glu Ser Pro Glu His Cys
      35      40      45
Ser Pro His His Thr Ala Leu Arg Gln Ala Ile Leu Cys Trp Gly Glu
      50      55      60
Leu Met Thr Leu Ala Thr Trp Val Gly Val Asn Leu Glu Asp Pro Ala
      65      70      75      80
Ser Arg Asp Leu Val Val Ser Tyr Val Asn Thr Asn Met Gly Leu Lys
      85      90      95
Phe Arg Gln Leu Leu Trp Phe His Ile Ser Cys Leu Thr Phe Gly Arg
      100      105      110
Glu Thr Val Ile Glu Tyr Leu Val Ser Phe Gly Val Trp Ile Arg Thr
      115      120      125
Pro Pro Ala Tyr Arg Pro Pro Asn Ala Pro Ile Leu Ser Thr Leu Pro
      130      135      140
Glu Thr Thr Val Val Arg Arg Arg Gly Arg Ser Pro Arg Arg Arg Thr
      145      150      155      160
Pro Ser Pro Arg Arg Arg Arg Ser Gln Ser Pro Arg Arg Arg Arg Ser
      165      170      175
Gln Ser Arg Glu Ser Gln Cys
      180
  
```

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 552 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

```

ATGGACATCG ACCCTTATAA AGAATTGGA GCTACTGTGG AGTTACTCTC GTTTTTGCCT      60
TCTGACTTCT TTCCTTCAGT ACGAGATCTT CTAGATACCG CCTCAGCTCT GTATCGGGAA      120
  
```

(2) INFORMATION FOR SEQ ID NO:40:

- (ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Thr Trp Val Gly Val Asn Leu Glu Asp Pro Ala Ser Arg Asp Leu Val
1 5 10 15
Val Ser Tyr Val Asn Thr Asn Met Gly
20 25

(2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Leu Leu Trp Phe His Ile Ser Cys Leu Thr Phe Gly Arg Glu Thr Val
1 5 10 15
Ile Glu Tyr Leu Val
20

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 16 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Val	Val	Ser	Tyr	Val	Asn	Thr	Asn	Met	Gly	Leu	Lys	Phe	Arg	Gln	Leu
1				5					10					15	

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Val	Ser	Phe	Gly	Val	Trp	Ile	Arg	Thr	Pro	Pro	Ala	Tyr	Arg	Pro	Pro
1				5					10					15	

Asn Ala Pro Ile Leu
20

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 35 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

GATCCCATGG ACATCGACCC TTATAAGAA TTTGG

35

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 44 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

GATCAAGCTT TTAACATTGA GATTCCCGAG ATTGAGATCT TCTG

44

(2) INFORMATION FOR SEQ ID NO:46:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 43 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:
GATCGAATTC ACTAGTTGGA AGATCCAGCG TCTAGAGACC TAG 43

(2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 41 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:
GATCGAATTC CTCGAGCTAG AGTCATTAGT TCCCCCAGC A 41

(2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:
GATTATCACT CGAGATGGCT TCCTGCTGCT GCAG 34

(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 40 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
GATCGAATTC AGCGCTCAAG CTCTGGAGGA AATCCACCAG 40

(2) INFORMATION FOR SEQ ID NO:50:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

Leu Asp Gly Cys Leu Leu Leu Gln Met Asp Phe Gly Phe Pro Lys His
1 5 10 15
Leu Leu Val Asp Phe Leu Gln Ser Leu Ser
20 25

(2) INFORMATION FOR SEQ ID NO:51:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 51 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

GGCCGCAACG TTTTACTAGC TCAGGCTCTG CAGGAAATCC ACCAGCAGGT G 51

(2) INFORMATION FOR SEQ ID NO:52:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 41 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

GGCCGCCCAT GGCCTGTCCC AAAGGCGCCT CCTACGAGGC T 41

Claims

1. A process for producing antibodies to
cholesteryl ester transfer protein (CETP) in a mammal
5 that comprises the steps of:

(a) immunizing said mammal with an
inoculum containing a vehicle in which is dissolved or
dispersed a recombinant DNA molecule comprising a DNA
sequence that contains (i) a sequence encoding a CETP
10 immunogen linked to (ii) a promoter sequence that
controls the expression of said CETP immunogen DNA
sequence in said mammal, said CETP immunogen being an
immunogenic polypeptide having a CETP amino acid residue
sequence, said immunization providing an amount of said
15 recombinant DNA molecule sufficient to induce antibodies
to CETP; and

(b) maintaining said immunized mammal for
a time period sufficient for the production of
antibodies that bind to CETP.
20

2. The process of claim 1 wherein the blood
of said mammal contains CETP.

3. A process for increasing the
25 concentration of HDL cholesterol in the blood of a
mammal whose blood contains cholesteryl ester transfer
protein (CETP) that comprises the steps of:

(a) immunizing said mammal with an
inoculum containing a vehicle in which is dissolved or
dispersed a recombinant DNA molecule comprising a DNA
30 sequence that contains (i) a sequence encoding a CETP
immunogen linked to (ii) a promoter sequence that
controls the expression of said CETP immunogen DNA
sequence in said mammal, said CETP immunogen being an
immunogenic polypeptide having a CETP amino acid residue
35

sequence, said immunization providing an amount of said recombinant DNA molecule sufficient to induce antibodies to CETP; and

5 (b) maintaining said immunized mammal for a time period sufficient for said CETP immunogen to be expressed and for the production of antibodies that bind to CETP and lessen the transfer of cholesteryl esters from HDL.

10 4. The process according to claim 3 wherein said immunizing step is repeated.

15 5. The process according to claim 3 wherein said immunizing step is repeated at intervals of about 3 to about 6 months until the HDL cholesterol value in the blood of said mammal is increased by about 10 percent or more relative to the HDL cholesterol value prior to said first immunization step.

20 6. The process according to claim 3 wherein said recombinant DNA molecule encodes human CETP as said immunogenic polypeptide.

25 7. The process according to claim 3 wherein said recombinant DNA molecule encodes rabbit CETP as said immunogenic polypeptide.

30 8. The process according to claim 3 wherein said encoded CETP immunogen comprises an immunogenic polypeptide fused to an exogenous antigenic carrier polypeptide.

9. The process according to claim 8 wherein said exogenous antigenic carrier polypeptide is selected

from the group consisting of hepatitis B core protein, tetanus toxoid, and diphtheria toxoid.

5 10. The process according to claim 9 wherein said recombinant DNA molecule encodes a fusion protein in which said exogenous antigenic carrier is fused to the carboxy-terminus of said immunogenic polypeptide.

10 11. The process according to claim 8 wherein the carboxy-terminus of said encoded exogenous antigenic carrier is fused to the amino-terminus of said encoded immunogenic polypeptide.

15 12. The process according to claim 8 wherein said encoded exogenous antigenic carrier is fused to both the amino-terminus and carboxy-terminus of said encoded immunogenic polypeptide.

20 13. The process according to claim 12 wherein said encoded fusion protein is comprised of an immunogenic polypeptide having a length of about 10 to about 30 amino acid residues that are fused to an amino-terminal flanking sequence and a carboxy-terminal flanking sequence, wherein

25 (a) said amino-terminal flanking sequence consists essentially of about 10 to about 20 amino acid residues having an amino acid residue sequence of the hepatitis B core protein (HBcAg) from about position 1 to about position 35, and said carboxy-terminal sequence consists essentially of about 120 to
30 about 160 amino acid residues having an amino acid residue sequence of HBcAg from about position 10 about position 183, or

35 (b) said amino-terminal flanking sequence consists essentially of about 70 to about 90 residues

having the amino acid residue sequence of HBcAg from about position 1 to about position 90, and said carboxy-terminal flanking sequence consists essentially of about 65 to about 85 amino acid residues having the amino acid residue sequence of HBcAg from about position 80 to about position 183.

14. The process according to claim 13 wherein the number of amino acid residues present in said encoded immunogenic polypeptide is about equal in number to the number of amino acid residues absent from said HBcAg amino acid residue sequence between the carboxy-terminal residue position of said amino-terminal flanking sequence and the amino-terminal residue of said carboxy-terminal flanking sequence.

15. The process according to claim 3 wherein said encoded immunogenic polypeptide has the amino acid residue sequence of SEQ ID NOs:29 or 50.

16. The process according to claim 3 wherein said immunization is carried out by injecting said inoculum into muscle or skin of said mammal.

17. An inoculum that comprises a recombinant DNA molecule comprising a DNA sequence that contains (i) a sequence encoding a CETP immunogen linked to (ii) a promoter sequence that controls the expression of said CETP immunogen DNA sequence in a mammal, said recombinant DNA molecule being dissolved or dispersed in an effective amount in a vehicle.

18. The inoculum of claim 17 wherein the concentration of said DNA encoding said CETP immunogen is about 0.05 μ g/ml to about 20 mg/ml.

19. The inoculum of claim 17 wherein said vehicle is phosphate-buffered saline.

5 20. The inoculum of claim 17 wherein said
vehicle is isotonic sucrose.

21. The inoculum of claim 17 wherein said DNA is complexed with liposomes.

10

Abstract of the Disclosure

5 A process for inducing the production of
antibodies that bind to cholesteryl ester transfer
protein (CETP) is disclosed. That process comprises the
steps of: (a) immunizing a mammal with an inoculum
containing a recombinant DNA molecule that comprises a
DNA sequence that contains (i) a sequence encoding a
10 CETP immunogen that is linked to (ii) a promoter
sequence that controls expression of the CETP immunogen,
the recombinant DNA molecule being dissolved or
dispersed in a vehicle; and (b) maintaining the
immunized mammal for a time period sufficient to induce
15 the production of antibodies that bind to CETP, and
preferably lessen the transfer of cholesteryl esters
from HDL where the blood of the mammal itself contains
CETP. Immunogens, inocula, DNA segments, and recombinant
DNA molecule vectors useful for carrying out the
20 invention are also disclosed.

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare:

That my residence, post office address and citizenship are as stated below next to my name.

That I verily believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: AN IMMUNOLOGICAL PROCESS AND CONSTRUCTS FOR INCREASING THE HDL CHOLESTEROL CONCENTRATION BY DNA VACCINATION

the specification of which (check one)

- ☐ is attached hereto.
- ☒ was filed on September 19, 1997 as
Application Serial No. 08/934,367
and was amended on _____
(if applicable)

That I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

That I acknowledge the duty to disclose information known to be material to patentability of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

That I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate on this invention having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No

That I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

United States Application(s)		
<u>08/785,997</u>	<u>January 21, 1997</u>	<u>Pending</u>
(Application Serial No.)	(Filing Date)	(Status)-(Patented, pending, abandoned)
<u>08/788,882</u>	<u>January 21, 1997</u>	<u>Pending</u>
(Application Serial No.)	(Filing Date)	(Status)-(Patented, pending, abandoned)
_____	_____	_____
(Application Serial No.)	(Filing Date)	(Status)-(Patented, pending, abandoned)

That all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

I hereby appoint the following attorneys, with full power of substitution and revocation, to prosecute this application and to transact all business in the United States Patent and Trademark Office connected therewith and request that all correspondence and telephone calls in respect to this application be directed to WELSH & KATZ, LTD., 120 South Riverside Plaza, 22nd Floor, Chicago, Illinois 60606, Telephone No. (312) 655-1500:

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Abstract

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